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(54) **Starburst conjugates.**

(57) **Starburst conjugates which are composed of at least one dendrimer in association with at least one unit of a carried agricultural, pharmaceutical, or other material have been prepared. These conjugates have particularly advantageous properties due to the unique characteristics of the dendrimer.**

STARBURST CONJUGATES

Field of the Invention

The present invention concerns the use of dense star polymers as carriers for agricultural, pharmaceutical, and other materials.

5 Background of the Invention

In recent years polymers referred to as dense star polymers or starburst polymers have been developed. It has been found that the size, shape and properties of these dense star polymers or starburst
10 polymers can be molecularly tailored to meet specialized end uses. Starburst polymers have significant advantages which can provide a means for the delivery of high concentrations of carried material per unit of polymer, controlled delivery, targeted delivery
15 and/or multiple species delivery or use.

Summary of the Invention

In its broadest aspect, the present invention
20 is directed to polymer conjugate materials comprising dense star polymers or starburst polymers associated with desired materials (hereinafter these polymer conjugates will frequently be referred to as "starburst

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conjugates" or "conjugates"), process for preparing these conjugates, compositions containing the conjugates, and methods of using the conjugates and compositions.

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The conjugates of the present invention are suitable for use in a variety of applications where specific delivery is desired, and are particularly suited for the delivery of biologically active agents.

10 In a preferred embodiment of the present invention, the starburst conjugates are comprised of one or more starburst polymers associated with one or more bioactive agents.

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The starburst conjugates offer significant benefits over other carriers known in the art due to the advantageous properties of the dense starburst polymers. Starburst polymers exhibit molecular

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architecture characterized by regular dendritic branching with radial symmetry. These radially symmetrical molecules are referred to as possessing

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"starburst topology". These polymers are made in a manner which can provide concentric dendritic tiers around an initiator core. The starburst topology is achieved by the ordered assembly of organic repeating units in concentric, dendritic tiers around an

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initiator core; this is accomplished by introducing multiplicity and self-replication (within each tier) in a geometrically progressive fashion through a number of molecular generations. The resulting highly

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functionalized molecules have been termed "dendrimers" in deference to their branched (tree-like) structure as well as their oligomeric nature. Thus, the terms starburst oligomer and starburst dendrimer are encompassed within the term starburst polymer.

Topological polymers, with size and shape controlled domains, are dendrimers that are covalently bridged through their reactive terminal groups, which are referred to as starburst "bridged dendrimers." The
5 term bridged dendrimer is also encompassed within the term "starburst polymer".

Brief Description of the Drawings

The following description of the figures aid in
10 understanding the present invention.

Figure 1 depicts various generations of starburst dendrimers.

15 Figure 2A depicts a dendrimer having unsymmetrical (unequal) branch junctures.

Figure 2B depicts a dendrimer having symmetrical (equal) branch junctures.

20 Figure 3 depicts dendrimer sizes relative to antibody dimensions.

Figure 4 shows carbon-13 spin lattice relaxation times (T₁) for aspirin incorporated into various dendrimer
25 generations, Example 1.

Figure 5 shows the results of the dynamic analysis of Example 2.

30 Figure 6 shows the influence of generation 6.5 dendrimer on the dialysis rate of pseudoephedrine at pH 9.5 from Example 2.

Figure 7 shows the effect of dendrimer hydrolysis on
35 the permeability of pseudoephedrine of Example 3.

Example 8 shows the comparison of percent salicylic acid released into the receptor compartment in the presence of starburst polymer (Gen = 4.0) at pH 5.0 and 6.65 with salicylic acid control, Example 4.

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Figure 9 shows the comparison of percent salicylic acid lost from donor compartment with starburst polymer (Gen = 4.0) in receptor compartment at pH 8.0 to salicylic acid content, Example 4.

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Figure 10 shows the comparison of percent salicylic acid lost from donor compartment in presence of starburst polymer (Gen = 4.5) to salicylic acid control, Example 4.

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Figure 11 shows carbon-13 spin lattice relaxation times (T_1) for 2,4-D incorporated into various dendrimer generations, Example 14.

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The starburst polymers are illustrated by Figure 1 wherein (I) represents an initiator core (in this figure a tri-functional initiator core shown by the far left drawing,); Z represents a terminal group, shown in the first instance by the second drawing from the left, referred to as a starbranched oligomer; A, B, C, D, and E represent particular molecular generations of starburst dendrimers; and $(A)_n$, $(B)_n$, $(C)_n$, $(D)_n$, and $(E)_n$ represent starburst bridged dendrimers.

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The starburst dendrimers are unimolecular assemblages that possess three distinguishing architectural features, namely, (a) an initiator core, (b) interior layers (generations, G) composed of repeating units, radially attached to the initiator core, and (c) an exterior surface of terminal functionality (i.e., terminal functional groups)

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attached to the outermost generation. The size and shape of the starburst dendrimer molecule and the functional groups present in the dendrimer molecule can be controlled by the choice of the initiator core, the number of generations (i.e., tiers) employed in creating the dendrimer, and the choice of the repeating units employed at each generation. Since the dendrimers can be readily isolated at any particular generation, a means is provided for obtaining dendrimers having desired properties.

The choice of the starburst dendrimer components affects the properties of the dendrimers. The initiator core type can affect the dendrimer shape, producing (depending on the choice of initiator core), for example, spheroid-shaped dendrimers, cylindrical or rod-shaped dendrimers, ellipsoid-shaped dendrimers, or mushroom-shaped dendrimers. Sequential building of generations (i.e., generation number and the size and nature of the repeating units) determines the dimensions of the dendrimers and the nature of their interior.

Because starburst dendrimers are branched polymers containing dendritic branches having functional groups distributed on the periphery of the branches, they can be prepared with a variety of properties. For example, starburst dendrimers, such as those depicted in Figure 2A and Figure 2B can have distinct properties due to the branch length. The dendrimer type shown in Figure 2A possesses unsymmetrical (unequal segment) branch junctures, exterior (i.e., surface) groups (represented by Z'), interior moieties (represented by Z) but much less internal void space. The dendrimer type shown in

Figure 2B possesses symmetrical (equal segment) branch
junctions with surface groups (represented by Z'), two
different interior moieties (represented respectively
by X and Z) with interior void space which varies as a
function of the generation (G). The dendrimers such
as those depicted in Figure 2B can be advanced through
enough generations to totally enclose and contain void
space, to give an entity with a predominantly hollow
interior and a highly congested surface.

Also, starburst dendrimers, when advanced
through sufficient generations exhibit "starburst dense
packing" where the surface of the dendrimer contains
sufficient terminal moieties such that the dendrimer
surface becomes congested and encloses void spaces
within the interior of the dendrimer. This congestion
can provide a molecular level barrier which can be used
to control diffusion of materials into or out of the
interior of the dendrimer.

Surface chemistry of the dendrimers can be
controlled in a predetermined fashion by selecting a
repeating unit which contains the desired chemical
functionality or by chemically modifying all or a
portion of the surface functionalities to create new
surface functionalities. These surfaces may either be
targeted toward specific sites or made to resist uptake
by particular organs or cells e.g. by reticuloendo-
thelial cells.

In an alternative use of the starburst
dendrimers, the dendrimers can themselves be linked
together to create polydendric moieties (starburst

"bridged dendrimers") which are also suitable as carriers.

5 In addition, the dendrimers can be prepared so as to have deviations from uniform branching in particular generations, thus providing a means of adding discontinuities (i.e., deviations from uniform branching at particular locations within the dendrimer) and different properties to the dendrimer.

10 The starburst polymers employed in the starburst conjugates of the present invention can be prepared according to methods known in the art, for example, U. S. Patent 4,587,329.

15 Dendrimers can be prepared having highly uniform size and shape and most importantly allow for a greater number of functional groups per unit of surface area of the dendrimer, and can have a greater number of functional groups per unit of molecular volume as compared to other polymers which have the same molecular weight, same core and monomeric components and same number of core branches as the starburst polymers. The increased functional group density of
20 the dense starburst polymers may allow a greater quantity of material to be carried per dendrimer. Since the number of functional groups on the dendrimers can be controlled on the surface and within the
25 interior, it also provides a means for controlling, for example, the amount of bioactive agent to be delivered per dendrimer. In a particularly preferred embodiment of the present invention, the starburst polymers, particularly the starburst dendrimers, are targeted
30 carriers of bioactive agents capable of delivering the
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bioactive agents to a particular target organism or to a particular determinant or locus in a target organism.

5 An analogy can be made between early generation starburst dendrimers (i.e. generation = 1-7) and classical spherical micelles. The dendrimer-micelles analogy was derived by comparing features which they had in common such as shape, size and surface.

10 Table I

	<u>Parameter</u>	<u>Regular Classical Micelles</u>	<u>Starburst Dendrimers</u>
	Shape	Spherical	Spherical
15	Size (diameter)	20-60Å	17-40Å
	Surface aggregation number	4-202	2=6-192 (Z is the number of surface groups) (generation = 2-7)
20	area/surface group (Å ²)	130-80Å ²	127-75Å ²

(1Å = 10⁻¹nm; 1Å² = 10⁻²nm²)

25 In Table I, the shape was verified by scanning transmission electron micrographs (STEM) microscopy and intrinsic viscosity (η) measurements. The size was verified by intrinsic viscosity [η] and size exclusion chromatography (SEC) measurements. The surface aggregation numbers were verified by titrimetry and high field NMR. The area/surface group was calculated from
30 SEC hydrodynamic measurements.

The first five generations of starburst polyamidoamine (PAMAM) dendrimers are microdomains
35 which very closely mimic classical spherical micelles in nearly every respect (i.e., shape, size, number of

surface groups, and area/surface groups). A major difference, however, is that they are covalently fixed and robust compared to the dynamic equilibrating nature of micelles. This difference is a significant
5 advantage when using these microdomains as encapsulation devices.

As further concentric generations are added beyond five, congestion of the surface occurs. This
10 congestion can lead to increased barrier characteristics at the surface and manifests itself as a smaller surface area per head (surface) group as shown in Table II.

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Table II
PAMAM Dendrimer Features vs. Generation

Generations	1	2	3	4	5	6	7	8	9
# of surface groups, Z	3	6	12	24	48	96	192	384	768
Molecular wt.	275	875	2411	5147	10,619	21,563	43,541	87,227	174,779
Diameter* measured SEC	10.4Å	15.8Å	22Å	31Å	40Å	53Å	67Å	76Å	88Å
Surface area per dendrimer	366Å ²	783Å ²	1519Å ²	3018Å ²	5024Å ²	8,820Å ²	14,096Å ²	18,136Å ²	36,083Å ²
Surface area per 2 group	122Å ²	131Å ²	127Å ²	126Å ²	104Å ²	92Å ²	73Å ²	47Å ²	32Å ²
Distance between 2 groups	12.4Å	12.8Å	12.7Å	12.6Å	11.5Å	10.8Å	9.8Å	7.75Å	6.28Å
Void Volume	311.6Å ³	1,470.2Å ³	4,737.9Å ³	11,427.0Å ³	---	---	---	---	---

Hydrodynamic diameters determined by size exclusion chromatography measurements calibrated against monodisperse ($\frac{M_w}{M_n} = 1.02$) polyethyleneoxide standards.

Å = 10⁻¹nm; 1Å² = 10⁻²nm²; 1Å³ = 10⁻³nm³.

For example, amine terminated generations 5.0, 6.0, 7.0, 8.0 and 9.0 have decreased surface areas of 104, 92, 73, 47 and 32\AA^2 per Z group, respectively. This characteristic corresponds to a transition from a
5 less congested micelle-like surface to a more congested bi-layer/monolayer barrier-like surface normally associated with vesicles (liposomes) or Langmuir-Blodgett type membranes.

10 If this surface congestion is occurring, the change in physical characteristics and morphology should be observed as the generations increase from the intermediate generation (6-8) to the more advanced
15 generations (9 or 10). The scanning transmission electron micrographs (STEM) for generations = 7.0, 8.0 and 9.0 were obtained after removing the methanol solvent from each of the samples to provide colorless, light yellow solid films. The morphological change
20 predicted occurred at the generation G=9.0 stage. The microdomains at generation = 9.0 measure about 33\AA in diameter and are surrounded by a colorless rim which is about 25\AA thick. Apparently methanolic solvent has been entrapped within the 25\AA outer membrane-like
25 barrier to provide the dark stained interior. Thus, at generation = 9.0, the starburst PAMAM is behaving topologically like a vesicle (liposome). However, this starburst is an order of magnitude smaller and very monodispersed compared to a liposome. Consequently,
30 the present dendrimers can be used to molecularly encapsulate solvent filled void spaces of as much diameter as about 33\AA (volume about $18,000\text{\AA}^3$) or more. These micelle sized prototypes appear to behave like a
35 covalently fixed liposome in this advanced generation stage. This behavior enables these prototypes to have

additional capability as carriers for, for example, non-chelating radionuclides in starburst antibody conjugates for the treatment of various mamalian diseases.

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Since the number of functional groups on the dendrimers can be controlled on the surface and within the interior, it also provides a means for controlling the amount of carried material to be delivered per
10 dendrimer. In one embodiment, the dendriers are targeted carriers of agents capable of delivering the carried material, for example, a bioactive agent, to, for example, a plant or pest or a particular determinant or locus in a target organism.

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Dendrimers suitable for use in the conjugates of the present invention include the dense star polymers or starburst polymers described in U. S. Patents 4,507,466, 4,558,120, 4,568,737 and 4,587,329.
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In particular, the present invention concerns a starburst conjugate which comprises at least one starburst polymer associated with at least one carried agricultural, pharmaceutical, or other material.
25 Starburst conjugates included within the scope of the present invention include those represented by the formula:

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$$(P)_x * (M)_y \quad (I)$$

wherein each P represents a dendrimer;

x represents an integer of 1 or greater;

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each M represents a unit (for example, a molecule, atom, ion, and/or other basic unit) of a carried material, said carried material can be the same carried material or a different carried material, preferably
5 the carried material is a bioactive agent;

y represents an integer of 1 or greater; and

* indicates that the carried material is associated
10 with the dendrimer.

Preferred starburst conjugates of formula (I) are those in which M is a drug, pesticide, radionuclide, chelant, chelated metal, toxin, antibody, antibody fragment,
15 antigen, signal generator, for example, fluorescing entities, signal reflector, for example, paramagnetic entities, or signal absorber, for example, electron beam opacifiers, fragrance, pheromones, or dyes. It is particularly preferred that $x=1$ and $y=2$ or more.

20 Also included are starburst conjugates of formula (I) wherein the starburst dendrimers are covalently linked together, starburst bridged dendrimers, optionally via linking groups, so as to form polydendric assemblages
25 (i.e., where $x>1$). Uses of these starburst bridged dendrimers include topical controlled release agents, radiation synovectomy, and others.

As used herein, "associated with" means that
30 the carried material(s) can be encapsulated or entrapped within the core of the dendrimer, dispersed partially or fully throughout the dendrimer, or attached or linked to the dendrimer, or any combination thereof. The association of the carried material(s)
35 and the dendrimer(s) may optionally employ connectors and/or spacers to facilitate the preparation or use of

the starburst conjugates. Suitable connecting groups are groups which link a targeting director (i.e., T) to the dendrimer (i.e., P) without significantly impairing the effectiveness of the director or the effectiveness of any other carried material(s) (i.e., M) present in the starburst conjugate. These connecting groups may be cleavable or non-cleavable and are typically used in order to avoid steric hindrance between the target director and the dendrimer, preferably the connecting groups are stable (i.e., non-cleavable). Since the size, shape and functional group density of the starburst dendrimers can be rigorously controlled, there are many ways in which the carried material can be associated with the dendrimer. For example, (a) there can be covalent, coulombic, hydrophobic, or chelation type association between the carried material(s) and entities, typically functional groups, located at or near the surface of the dendrimer; (b) there can be covalent, coulombic, hydrophobic, or chelation type association between the carried material(s) and moieties located within the interior of the dendrimer; (c) the dendrimer can be prepared to have an interior which is predominantly hollow allowing for physical entrapment of the carried materials within the interior (void volume), wherein the release of the carried material can optionally be controlled by congesting the surface of the dendrimer with diffusion controlling moieties; or (d) various combinations of the aforementioned phenomena can be employed.

Dendrimers, herein represented by "P", include the dense star polymers described in U. S. Patent 4,507,466, 4,558,120, 4,568,737 or 4,587,329.

In a preferred embodiment, the carried materials, herein represented by "M", are pharmaceutical materials. Such materials which are suitable for use in the starburst conjugates include any materials for in vivo or in vitro use for diagnostic or therapeutic treatment of mammals which can be associated with the dense star dendrimer without appreciably disturbing the physical integrity of the dendrimer, for example, drugs such as antibiotics, analgesics, hypertensives, cardiotonics, and the like such as acetaminaphen, acyclovir, alkeran, amikacin, ampicillin, aspirin, bisantrene, bleomycin, neocardioastatin, chloroambucil, chloramphenicol, cytarabine, daunomycin, doxorubicin, fluorouracil, gentamycin, ibuprofen, kanamycin, meprobamate, methotrexate, novantrone, nystatin, oncovin, phenobarbital, polymyxin, probucol, procarbazine, rifampin, streptomycin, spectinomycin, symmetrel, thioguanine, tobramycin, trimethoprim, valban; and toxins such as diphtheria toxin, gelonin, exotoxin A, abrin, modeccin, ricin, or toxic fragments thereof; metal ions such as the alkali and alkaline-earth metals; radionuclides such as those generated from actinides or lanthanides or other similar transition elements or from other elements, such as ^{67}Cu , ^{90}Y , ^{111}In , ^{131}I , ^{186}Re , ^{105}Rh , $^{99\text{m}}\text{Te}$, ^{67}Ga , ^{153}Sm , ^{159}Gd , ^{175}Yb , ^{177}Lu , ^{88}Y , ^{166}Ho , $^{115\text{m}}\text{In}$, ^{109}Pd , ^{82}Rb , ^{194}Ir , ^{140}Ba , ^{149}Pm , ^{199}Au , ^{140}La , and ^{188}Re ; signal generators such as fluorescing entities; signal reflectors such as paramagnetic entities, for example, ^{56}Fe , Gd , ^{55}Mn ; chelated metal such as any of the metals given, whether or not they are radioactive, when associated with a chelant; signal absorbers such as an electron beam opacifiers; antibodies including

monoclonal antibodies and anti-idiotypic antibodies; antibody fragments; hormones; biological response modifiers such as interleukins, interferons, viruses and viral fragments; diagnostic opacifiers; and
5 fluorescent moieties. Carried pharmaceutical materials include scavenging agents such as chelants, antigens, antibodies or any moieties capable of selectively scavenging therapeutic or diagnostic agents.

10 In another embodiment, the carried materials, herein represented by "M", are agricultural materials. Such materials which are suitable for use in the starburst conjugates include any materials for in vivo
15 or in vitro treatment, diagnosis, or application to plants or non-mammals (including microorganisms) which can be associated with the starburst dendrimer without appreciably disturbing the physical integrity of the dendrimer. For example, the carried materials can be
20 toxins such as diphtheria toxin, gelonin, exotoxin A, abrin, modeccin, ricin, or toxic fragments thereof; metal ions such as the alkali and alkaline earth metals; radionuclides such as those generated from actinides or lanthanides or other similar transition
25 elements or from other elements, such as ^{67}Cu , ^{90}Y , ^{111}In , ^{131}I , ^{186}Re , ^{105}Rh , $^{99\text{m}}\text{Te}$, ^{67}Ga , ^{153}Sm , ^{159}Gd , ^{175}Yb , ^{177}Lu , ^{88}Y , ^{166}Ho , $^{115\text{m}}\text{In}$, ^{109}Pd , ^{82}Rb , ^{194}Ir , ^{140}Ba , ^{149}Pm , ^{199}Au , ^{140}La , and ^{188}Re ; signal
30 generators such as fluorescing entities; signal reflectors such as paramagnetic entities; signal absorbers such as electron beam opacifiers; hormones; biological response modifiers such as interleukins, interferons, viruses and viral fragments; pesticides,
35 including antimicrobials, algicides, arithelmetics, acaricides, II insecticides, attractants, repellants,

herbicides and/or fungicides such as acephate, acifluorfen, alachlor, atrazine, benomyl, bentazon, captan, carbofuran, chloropicrin, chlopyrifos, chlorsulfuron cyanazine, cyhexatin, cypermethrin, 5 2,4-dichlorophenoxyacetic acid, dalapon, dicamba, diclofop methyl, diflubenuron, dinoseb, endothall, ferbam, fluazifop, glyphosate, haloxyfop, malathion, naptalam, pendimethalin, permethrin, picloram, propachlor, propanil, sethoxydin, temephos, terbufos, 10 trifluralin, triforine, zineb, and the like. Carried agricultural materials include scavenging agents such as chelants, chelated metal (whether or not they are radioactive) or any moieties capable of selectively scavenging therapeutic or diagnostic agents. 15

In a further embodiment, the carried materials, herein represented by "M", which are suitable for use in the starburst conjugates include any materials other than agricultural or pharmaceutical materials which can 20 be associated with the starburst dendrimer without appreciably disturbing the physical integrity of the dendrimer, for example, metal ions such as the alkali and alkaline-earth metals; signal generators such as fluroescing entities; signal reflectors such as 25 paramagnetic entities; signal absorbers such as an electron beam opacifiers; pheromone moieties; fragrance moieties; dye moieties; and the like. Carried materials include scavenging agents such as 30 chelants or any moieties capable of selectively scavenging a variety of agents.

Preferably the carried materials are bioactive agents. As used herein, "bioactive" refers to an 35 active entity such as a molecule, atom, ion and/or other entity which is capable of detecting,

identifying, inhibiting, treating, catalyzing, controlling, killing, enhancing or modifying a targeted entity such as a protein, glycoprotein, lipoprotein, lipid, a targeted cell, a targeted organ, a targeted
5 organism [for example, a microorganism, plant or animal (including mammals such as humans)] or other targeted moiety.

The starburst conjugates of formula (I) are
10 prepared by reacting P with M, usually in a suitable solvent, at a temperature which facilitates the association of the carried material (M) with the starburst dendrimer (P).

15 Suitable solvents are solvents in which P and M are at least partially miscible and inert to the formation of the conjugate. If P and M are at least partially miscible with each other, no solvent may be required. When desired, mixtures of suitable solvents
20 can be utilized. Examples of such suitable solvents are water, methanol, ethanol, chloroform, acetonitrile, toluene, dimethylsulfoxide and dimethylformamide.

25 The reaction conditions for the formation of the starburst conjugate of formula (I) depend upon the particular dendrimer (P), the carried material (M), and the nature of the bond (*) formed. For example, if P is the PEI (polyethyleneimine) starburst dendrimer with a methylene carboxylate surface, M is a radionuclide,
30 e.g. yttrium, then the reaction is conducted at room temperature in water. However, if P is an ester terminated PAMAM starburst dendrimer, M is aspirin, then the reaction is conducted at room temperature in
35 chloroform. Typically, the temperature can range from room temperature to reflux. The selection of the

particular solvent and temperature will be apparent to one skilled in the art.

5 The ratio of M:P will depend on the size of the dendrimer and the amount of carried material. For example, the molar ratio (ratio of moles) of any ionic M to P usually is 0.1-1,000:1, preferably 1-50:1, and more preferably 2-6:1. The weight ratio of any drug, pesticide, organic or toxin M to P usually is 0.1-5:1, 10 and preferably 0.5-3:1.

 When M is a radionuclide, there are three ways the starburst conjugate can be prepared, namely: (1) P can be used as a chelant. For example, a methylene- 15 carboxylate surface PEI or PAMAM will chelate a metal such as yttrium or indium. (2) A chelate can be covalently bonded to P. For example, an amine terminated PEI starburst dendrimer can be reacted with 1-(p-isothiocyanatobenzyl)diethylenetriaminepenta- 20 acetic acid and then chelated, or a complex such as rhodium chloride chelated with isothiocyanatobenzyl-2,3,2-tet can be reacted. (3) A prechelated radionuclide can be associated with P by hydrophobic or 25 ionic interaction.

 Other starburst conjugates, which are particularly preferred for use with pharmaceutical materials, are those conjugates which contain a target 30 director (herein designated as "T") and which are represented by the formula:



wherein

each T represents a target director;

5 e represents an integer of 1 or greater; and

P, x, *, M, and y are as previously defined herein.

Preferred among the starburst conjugates of formula
10 (II) are those in which M is a drug, pesticide,
radionuclide, chelant, toxin, signal generator, signal
reflector, or signal absorber. Also preferred
conjugates are those conjugates in which e=1 or 2; and
those in which x=1 and y=2 or more. Particularly
15 preferred conjugates are those in which x=1, e=2, y=2
or more and M and T are associated with the polymer via
the same or different connectors.

The starburst conjugates of formula (II) are
20 prepared either by forming T*P and then adding M or by
forming P*M and then adding T. Either reaction scheme
is conducted at temperatures which are not detrimental
to the particular conjugate component and in the
presence of a suitable solvent when required. To
25 control pH, buffers or addition of suitable acid or
base is used. The reaction conditions are dependent on
the type of association formed (*), the starburst
dendrimer used (P), the carried material (M), and the
30 target director (T). For example, when T is a
monoclonal antibody and M is a radionuclide, the T*P
association is done through a function group such as an
isothiocyanate in water or in water with an organic
modifier such as acetonitrile or dimethylformamide.
35 Usually, the conjugation is done in a buffer at pH
7-10, preferably pH 8.5-9.5. The formed conjugate is

then chelated with a radionuclide such as yttrium acetate, preferably at room temperature. Alternatively, P and M can be chelated, usually in water, before conjugation to T. The conjugation with T is
5 carried out in a suitable buffer.

The ratio of T:P is preferably 1:1, especially when T is an antibody or fragment. The ratio of M:P will be as before.

10 Target directors capable of targeting the starburst conjugates are entities which when used in the starburst conjugates of the present invention result in at least a portion of the starburst
15 conjugates being delivered to a desired target (for example, a protein, glycoprotein, lipoprotein, lipid, a targeted cell, a targeted organ, a targeted organism or other targeted moiety) and include antibodies, preferably monoclonal antibodies, antibody fragments
20 such as Fab, Fab', F(ab')₂ fragments or any other antibody fragments having the requisite target specificity, hormones, biological response modifiers; epitopes; chemical functionalities exhibiting target
25 specificity; and the like.

The antibodies or antibody fragments which may be used in preferred starburst conjugates described herein can be prepared by techniques well known in the
30 art. Highly specific monoclonal antibodies can be produced by hybridization techniques well known in the art, see, for example, Kohler and Milstein (1975, Nature 256:495-497; and 1976, Eur. J. Immunol. 6:511-519). Such antibodies normally have a highly specific
35 reactivity.

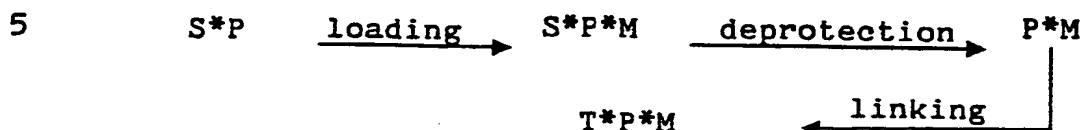
In the antibody targeted starburst conjugates, antibodies directed against any antigen or hapten may be used. Although conventional polyclonal antibodies may be used, monoclonal antibodies offer several
5 advantages. Each monoclonal antibody is highly specific for a single epitope. In addition, large amounts of each monoclonal antibody can be produced. Antibodies used in the present invention may be directed against, for example, tumors, bacteria, fungi,
10 viruses, parasites, mycoplasma, differentiation and other cell membrane antigens, pathogen surface antigens, toxins, enzymes, allergens, drugs and any biologically active molecules. For a more complete
15 list of antigens see U. S. Patent 4,193,983.

It may be desirable to connect more antibodies or fragments to the dendrimer, and in particular instances to connect antibodies of different
20 specificities. For example, a bifunctional conjugate which has the ability to localize and bind to a tumor and then scavenge circulating cytotoxic, diagnostic, or biostatic compounds can be designed.

25 In the absence of a target director (or in the presence of a target director if desired), due to the number of functional groups which can be located at or near the surface of the dendrimer, all or a substantial portion of such functional groups can be made anionic,
30 cationic, hydrophobic or hydrophilic to effectively aid delivery of the starburst conjugate to a desired target of the opposite charge or to a hydrophobic or hydrophilic compatible target.

35 Preparation of the conjugates of formula (II) using a P with a protected handle (S) is also intended

as a process to prepare the conjugates of formula (II). The reaction scheme is shown below:



10 where

S*P represents the protected dendrimer;

S*P*M represents the protected dendrimer conjugated with M;

15 P*M represents the dendrimer conjugated with M (starburst conjugate);

T*P*M. represents the starburst conjugate linked to the target director.

20 Suitable solvents can be employed which do not effect P*M. For example when S is t-butoxycarbamate, S can be removed by aqueous acid.

25 Also preferred when the carried materials are pharmaceutical materials are starburst conjugates in which the polymer is associated directly, or via connectors; these starburst conjugates are represented by the formula:

30

$$[(T)_e - (C')_f]_g * (P)_x * [(C'')_h - (M)_y]_k \quad (III)$$

wherein

35

each C' represents the same or different connecting group;

5 each C'' represents the same or different connecting group;

g and k each individually represent an integer of 1 or greater;

10 f and h each individually represent an integer of 0 or greater;

- indicates a covalent bond in instances where a connecting group is present; and

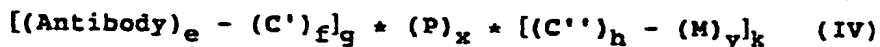
15 P, x, *, M, y, T, and e are as previously defined herein.

Preferred among the starburst conjugates of formula (III) are those in which M is a radionuclide, drug, 20 toxin, signal generator, signal reflector or signal absorber. Also preferred are those conjugates in which x=1. Particularly preferred conjugates are those in which x, e, f, h, and y are each 1, and g is 1 or more and k is 2 or more. Most preferred are those 25 conjugates in which x, e, f, h, y and g are each 1, and k is 2 or more. Also particularly preferred are those starburst conjugates in which M represents a bioactive agent such as a radionuclide, drug, or toxin.

30 Suitable connecting groups which are represented by C' are groups which link the carried pharmaceutical material to the dendrimer without significantly impairing the effectiveness of the 35 carried pharmaceutical material or the effectiveness of the target director(s) present in the starburst conju-

gate. These connectors must be stable (i.e., non-cleavable) or cleavable depending upon the mode of activity of the carried pharmaceutical material and are typically used in order to avoid stearic hindrance
 5 between the carried pharmaceutical material and the polymer.

Most preferred are conjugates in which the dendrimer is associated directly, or via connecting
 10 group(s), to one antibody or antibody fragment. The polymer in these preferred conjugates may, in addition, be optionally associated either directly, or via connecting group(s), to one or more other carried
 15 materials, preferably a radioisotope. Such starburst conjugates are represented by the formula:



20 wherein

each Antibody represents an antibody or antibody fragment capable of interacting with a desired epitope;

25 - indicates a covalent or coulombic bond in instances where a connecting group is present; and

P, x, *, M, T, e, y, C', C'', g, k, f, and h are as previously defined herein.

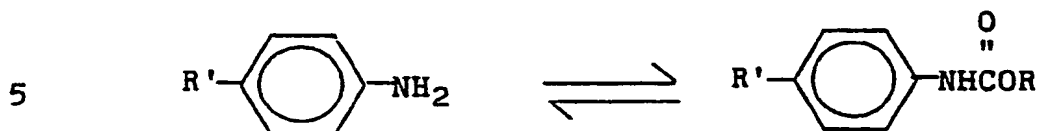
30 For the above synthesis of starburst dendrimers (P) which have a functional group available for linking (C' or C'') with a targeting director (T), the preferred process requires that the reactive functionality be
 35 protected as a synthetic precursor. This protection is preferred because it enables the synthesis of dendrimer

or conjugates of very high quality. This process allows for the chemical binding of a unit of carried pharmaceutical material (M) to the terminal functional groups of the starburst dendrimer (P) in ways which would otherwise result also in reaction with a linking functional group, thus making it impossible to attach to the targeting director (T). Subsequent deprotection or synthetic conversion into the desired linking functional group thus enables the starburst conjugate to be linked to the targeting director.

One of the preferred "functional groups for linking" (hereafter referred to as a "handle") is an aniline moiety. This group is preferred because it can be used directly for linking to the targeting director, or it can be readily modified to other functional groups suitable for reaction with the targeting director, e.g. isothiocyanate, isocyanate, semithiocarbazide, semicarbazide, bromoacetamide, iodoacetamide, and maleimide. The aniline moiety is also preferred as a handle for linking with the targeting directors because it can be readily protected for use in starburst dendrimer synthesis, or the nitro group can be used as a precursor which can be converted into the desired amino function at the end of the synthesis.

There are a number of protecting groups which are suitable for protecting the anilino amino functionality during starburst dendrimer synthesis. (See Theodora W. Green, Protective Groups In Organic Synthesis., Pub. John Wiley & Son, New York, 1981). A preferred class of protecting groups are the carbamates shown below.

-27-



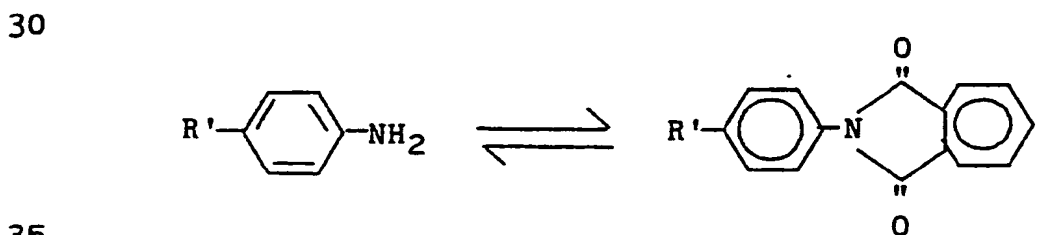
Many carbamates have been used for protection of
amines. The most preferred carbamates for starburst
dendrimer synthesis is the t-butoxycarbamate,
10 $\text{R} = \text{-C(CH}_3)_3$. Deprotection is achieved by mild acid
hydrolysis. Also preferred is the benzylcarbamate
protecting group,



which is preferred when the dendrimer is susceptible to
acid hydrolysis. Deprotection is achieved by catalytic
20 hydrogenation. Also preferred is the 9-fluorenyl-
methylcarbamate,



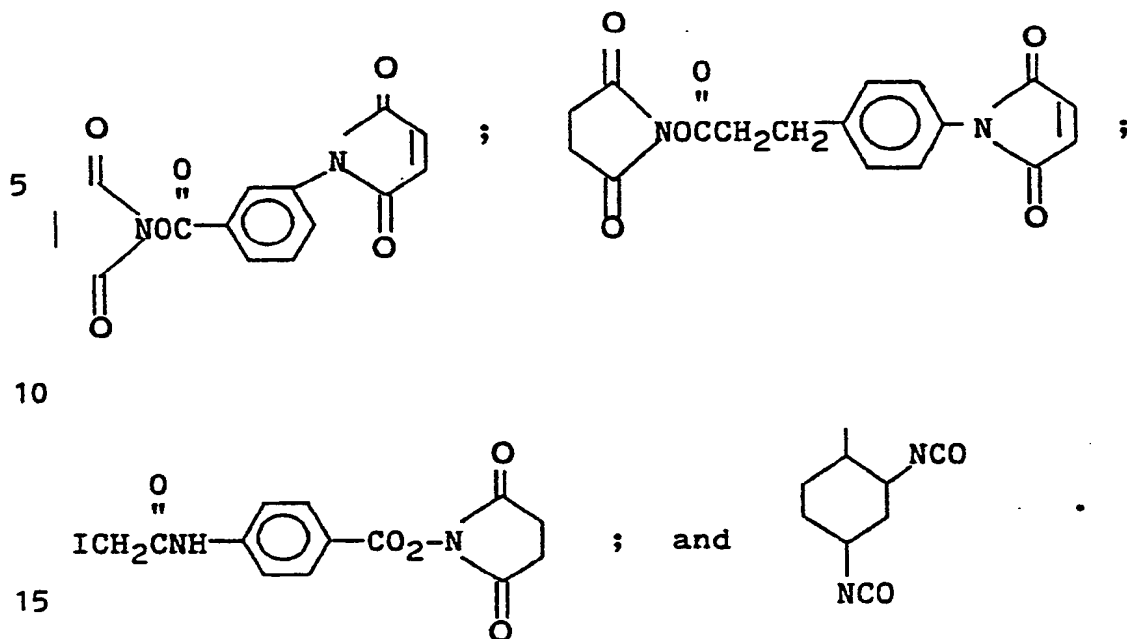
The phthalimide protecting group is also a
preferred example,



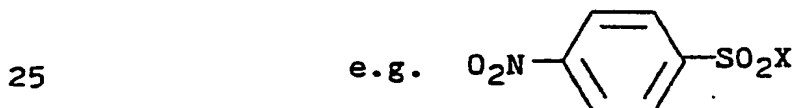
Other protecting groups used for amines which are well known in the literature could also be used in this synthetic scheme. The above preferences are given as illustrative examples only but are not the only carbamates which can be used. Any carbamate which is stable under the reaction conditions and can be removed without altering the integrity of the starburst dendrimer can be employed.

The above process can introduce an aminophenyl functionality into any agent containing an amino group which is then conjugated with some bioactive agent, e.g. monoclonal antibody or enzyme. The agent can be conjugated by oxidative coupling to carbohydrates on the bioactive agent, e.g. antibody. The aminophenyl group also can be converted into an isothiocyanate or isocyanate for subsequent reaction with the pendant amino groups of lysine residues on the bioactive agent.

The process involves the reaction of an activated aryl halide, e.g. 4-nitrofluorobenzene, with an amino-function; on the agent for conjugation, e.g. starburst polyethyleneimines (PEI), and subsequent catalytic hydrogenation of the nitro group to the aniline functionality for subsequent conjugation. It is particularly useful for agents, e.g. polyamines, which need further modification prior to use, due to the relative chemical inertness of the nitrophenyl functionality to all non-reducing reaction conditions. The more common bifunctional linking agents, e.g. active esters diisocyanates, which are reactive under a large number of reaction conditions and which would render them unusable for conjugation are:



20 The invention also includes the use of nitrosubstituted arylsulphonyl halides to give sulphonamides,



30 The advantage of this process over known processes of introducing an aminophenyl group for conjugation is that it takes place at a late stage of the synthesis. Gansow et al., U.S. Patent 4,472,509, in his process introduced the nitrophenyl group at the first step of a long synthetic procedure, thereby having limitations on the chemistry available.

35 This process also introduces a handle which is clearly differentiable from the remainder of the

molecule. Manabe et al., disclosed that the ring opening of succinic anhydride by residual amines gave a coupling group through which conjugation to an antibody was possible. This method however gave no means of
5 differentiating between any unchelated sites on the polymer, since the chelating groups were the same as the linking group.

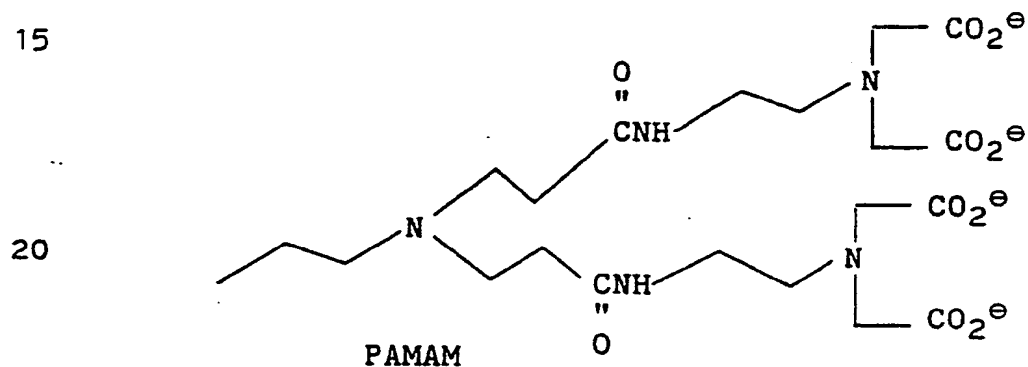
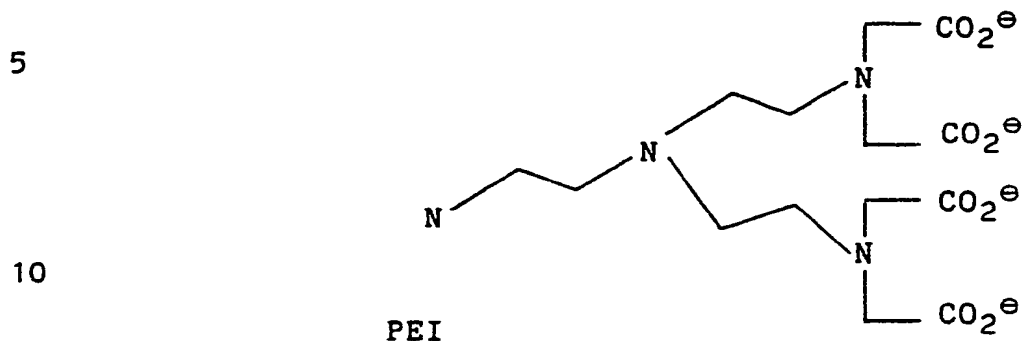
The present process also provides for direct
10 synthesis of lanthanides with starburst dendrimers, preferably by PEI acetate dendrimer. In contrast, Denkewalte, U.S. Patent 4,289,872, states that just putting acetates on the surface works. However, the present reaction shows that PEI acetate, works much
15 better than PAMAM, i.e., surface of iminodiacetates is only part of the story, the nature of the backbone, and branching is important also. The PEI acetate has better chelating properties than the PAMAM acetate.

20

25

30

35



25 Preferred among the starburst conjugates of formula (IV) are those in which M is a radionuclide, drug, toxin, signal generator, signal reflector or signal absorber. Also preferred are those conjugates

30 in which $x=1$. Particularly preferred are those conjugates in which x , e , f , h , and y are each 1, and g is 1 or more and k is each individually 2 or more. Most preferred are those conjugates in which x , e , f , h , y , and g are each 1, and k is 2 or more. Also

35 particularly preferred are those starburst conjugates in which "Antibody" represents a monoclonal antibody or

an epitope binding fragment thereof; and especially preferred are those in which M represents a bioactive agent such as a radionuclide, drug, or toxin.

5 The starburst conjugates can be used for a variety of in vitro or in vivo diagnostic applications such as radioimmunoassays, electron microscopy, enzyme linked immunosorbent assays, nuclear magnetic resonance spectroscopy, contrast imaging, and immunoscintigraphy, 10 in analytical applications, in therapeutic applications as a carrier of antibiotics, radionuclides, drugs or other agents suitable for use in the treatment of disease states such as cancer, autoimmune diseases, 15 central nervous system disorders, infectious diseases, and cardiac disorders; in biological control applications as a means of delivering pesticides such as herbicides, fungicides, repellants, attractants, antimicrobials or other toxins; or used as starting 20 materials for making other useful agents.

 The present invention is also directed to starburst conjugate compositions in which the starburst conjugates are formulated with other suitable vehicles. 25 The starburst conjugate compositions may optionally contain other active ingredients, additives and/or diluents.

 In the agricultural materials embodiment of the 30 invention, the starburst conjugates can be formulated with suitable vehicles useful in agriculture such as on crops, fallow land, or as pesticides, or in treatment of in vivo or in vitro testing of non-mammals. An agriculturally acceptable carrier or diluent which may 35 also be present with one or more starburst conjugates of the present invention includes those carriers or

diluents customarily used in granular formulations, emulsifiable concentrates, solutions, or suspensions such as, for example, toluene, xylene, benzene, phenol, water, methane, hydrocarbons, naphthalene and others.

5

The preferred starburst polymer for use in the starburst conjugates of the present invention is a polymer that can be described as a starburst polymer having at least one branch (hereinafter called a core
10 branch), preferably two or more branches, emanating from a core, said branch having at least one terminal group provided that (1) the ratio of terminal groups to the core branches is more than one, preferably two or greater, (2) the density of terminal groups per unit
15 volume in the polymer is at least 1.5 times that of an extended conventional star polymer having similar core and monomeric moieties and a comparable molecular weight and number of core branches, each of such
20 branches of the extended conventional star polymer bearing only one terminal group, and (3) a molecular volume that is no more than about 80 percent of the molecular volume of said extended conventional star polymer as determined by dimensional studies using
25 scaled Corey-Pauling molecular models. As used herein, the term "dense" as it modifies "star polymer" or "dendrimer" means that it has a smaller molecular volume than an extended conventional star polymer having the same molecular weight. The extended
30 conventional star polymer which is used as the base for comparison with the dense star polymer is one that has the same molecular weight, same core and monomeric components and same number of core branches as the
35 dense star polymer. By "extended" it is meant that the individual branches of the conventional star polymer

are extended or stretched to their maximum length, e.g., as such branches exist when the star polymer is completely solvated in an ideal solvent for the star polymer. In addition while the number of terminal
5 groups is greater for the dense star polymer molecule than in the conventional star polymer molecule, the chemical structure of the terminal groups is the same.

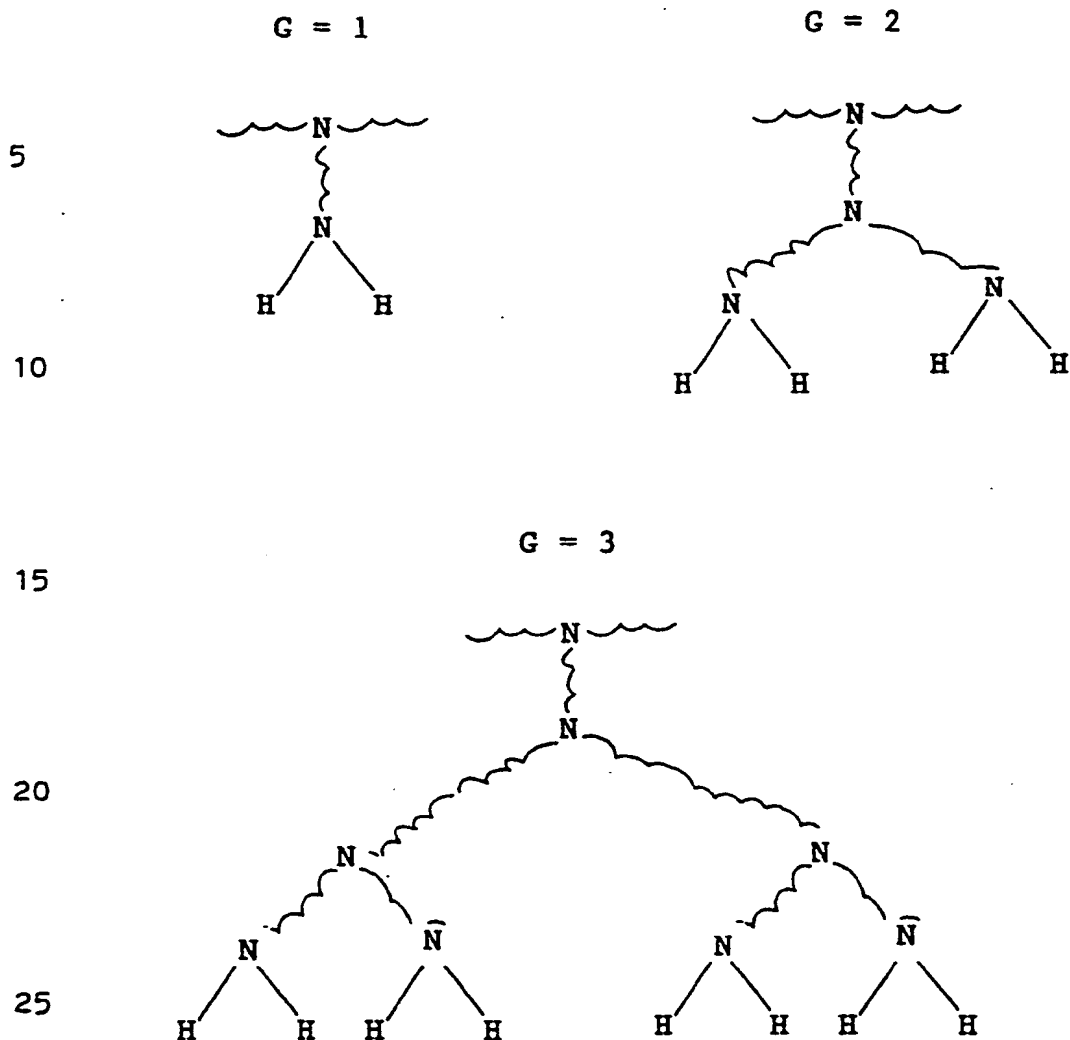
Dendrimers used in the conjugates of the
10 present invention can be prepared by processes known in the art. The above dendrimers, the various coreactants and core compounds, and process for their preparation can be as defined in U. S. Patent 4,587,329.

15 The dendrimers, for use in the conjugates of the present invention, can have terminal groups which are sufficiently reactive to undergo addition or substitution reactions. Examples of such terminal
20 groups include amino, hydroxy, mercapto, carboxy, alkenyl, allyl, vinyl, amido, halo, urea, oxiranyl, aziridinyl, oxazolinyll, imidazolinyll, sulfonato, phosphonato, isocyanato and isothiocyanato. The terminal groups can be modified to make them
25 biologically inert, for example, to make them non-immunogenic or to avoid non-specific uptake in the liver, spleen or other organ. The dendrimers differ from conventional star or star-branched polymers in that the dendrimers have a greater concentration of
30 terminal groups per unit of molecular volume than do conventional extended star polymers having an equivalent number of core branches and an equivalent core branch length. Thus, the density of terminal
35 groups per unit volume in the dendrimer usually is at least 1.5 times the density of terminal groups in the conventional extended star polymer, preferably at least

5 times, more preferably at least 10 times, most preferably from 15 to 50 times. The ratio of terminal groups per core branch in the dense polymer is preferably at least 2, more preferably at least 3, most preferably from 4 to 1024. Preferably, for a given polymer molecular weight, the molecular volume of the dense star polymer is less than 70 volume percent, more preferably from 16 to 60, most preferably from 7 to 50 volume percent of the molecular volume of the conventional extended star polymer.

Preferred dendrimers for use in the conjugates of the present invention are characterized as having a univalent or polyvalent core that is covalently bonded to dendritic branches. Such ordered branching can be illustrated by the following sequence wherein G indicates the number of generations:

-36-



Mathematically, the relationship between the number (#) of terminal groups on a dendritic branch and the number of generations of the branch can be represented as follows:

$$\# \text{ of terminal groups per dendritic branch} = \frac{N_r^G}{2}$$

wherein G is the number of generations and N_r is the repeating unit multiplicity which is at least 2 as in the case of amines. The total number of terminal groups in the dendrimer is determined by the following:

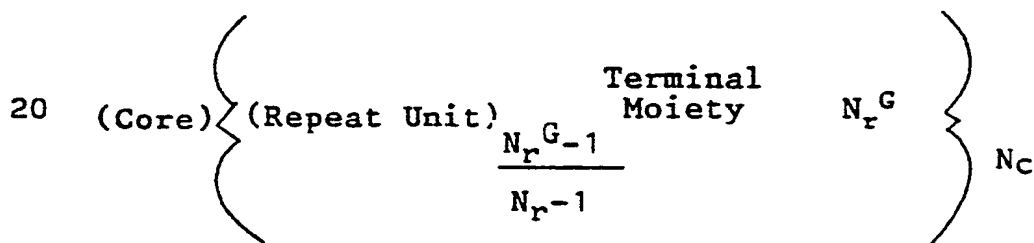
5

$$\# \text{ of terminal groups per dendrimer} = \frac{N_c N_r^G}{2}$$

10

wherein G and N_r are as defined before and N_c represents the valency (often called core functionality) of the core compound. Accordingly, the dendrimers of this invention can be represented in its component parts as follows:

15



25

wherein the Core, Terminal Moiety, G and N_c are as defined before and the Repeat Unit has a valency or functionality of $N_r + 1$ wherein N_r is as defined before.

30

A copolymeric dendrimer which is a preferred dendrimer for the purposes of this invention is a unique compound constructed of polyfunctional monomer units in a highly branched (dendritic) array. The dendrimer molecule is prepared from a polyfunctional initiator unit (core compound), polyfunctional

35

repeating units and terminal units which may be the same or different from the repeating units. The core compound is represented by the formula $\textcircled{\text{I}} (\text{Z}^{\text{C}})_{\text{Nc}}$ wherein $\textcircled{\text{I}}$ represents the core, Z^{C} represents the functional groups bonded to $\textcircled{\text{I}}$ and Nc represents the core functionality which is preferably 2 or more, most preferably 3 or more. Thus, the dendrimer molecule comprises a polyfunctional core, $\textcircled{\text{I}}$, bonded to a number (Nc) of functional groups, Z^{C} , each of which is connected to the monofunctional tail of a repeating unit, $\text{X}^1\text{Y}^1(\text{Z}^1)_{\text{N}^1}$, of the first generation and each of the Z groups of the repeating unit of one generation is bonded to a monofunctional tail of a repeating unit of the next generation until the terminal generation is reached.

In the dendrimer molecule, the repeating units are the same within a single generation, but may differ from generation to generation. In the repeating unit, $\text{X}^1\text{Y}^1(\text{Z}^1)_{\text{N}^1}$, X^1 represents the monofunctional tail of the first generation repeating unit, Y^1 represents the moiety constituting the first generation, Z^1 represents the functional group of the polyfunctional head of the repeating unit of the first generation and may be the same as or different from the functional groups of the core compound, $\textcircled{\text{I}} (\text{Z}^{\text{C}})_{\text{Nc}}$, or other generations; and N^1 is a number of 2 or more, most preferably 2, 3 or 4, which represents the multiplicity of the polyfunctional head of the repeating unit in the first generation. Generically, the repeating unit is represented by the formula $\text{X}^i\text{Y}^i(\text{Z}^i)_{\text{N}^i}$ wherein "i" represents the particular generation from the first to the t-1 generation. Thus, in the preferred dendrimer molecule, each Z^1 of the first generation repeating unit is

connected to an X^2 of a repeating unit of the second generation and so on through the generations such that each Z^i group for a repeating unit $X^i Y^i (Z^i)_{N^i}$ in generation number "i" is connected to the tail (X^{i+1}) of the repeating unit of the generation number "i+1".
 5 The final or terminal of a preferred dendrimer molecule comprises terminal units, $X^t Y^t (Z^t)_{N^t}$ wherein t represents terminal generation and X^t , Y^t , Z^t and N^t may be the same as or different from X^i , Y^i , Z^i and N^i
 10 except that there is no succeeding generation connected to the Z^t groups and N^t may be less than two, e.g., zero or one. Therefore, the preferred dendrimer has a molecular formula represented by

15

$$\left(\textcircled{I} (Z^c)_{N^c} \right) \left\{ \left(X^i Y^i (Z^i)_{N^i} \right)_{N^c \prod_{n=1}^{i-1} N^n} \right\} \left(X^t Y^t (Z^t)_{N^t} \right)_{N^c \prod_{n=1}^{t-1} N^n}$$

20

where i is 1 to t-1

wherein the symbols are as previously defined. The π
 25 function is the product of all the values between its defined limits. Thus

$$\prod_{n=1}^{i-1} N^n = (N^1)(N^2)(N^3)\dots(N^{i-2})(N^{i-1})$$

30

which is the number of repeat units, $X^i Y^i (Z^i)_{N^i}$, comprising the ith generation of one dendritic branch and when i is 1, then

35

$$\prod_{n=1}^0 = 1$$

In copolymeric dendrimers, the repeat unit for one generation differs from the repeat unit in at least one other generation. The preferred dendrimers are very symmetrical as illustrated in structural formulas described hereinafter. Preferred dendrimers may be converted to functionalized dendrimers by contact with another reagent. For example, conversion of hydroxyl in the terminal generation to ester by reaction with an acid chloride gives an ester terminally functionalized dendrimer. This functionalization need not be carried out to the theoretical maximum as defined by the number of available functional groups and, thus, a functionalized dendrimer may not have high symmetry or a precisely defined molecular formula as is the case with the preferred dendrimer.

In a homopolymeric dendrimer, all of the repeat units, $X^i Y^i (Z^i)_{N^i}$, are identical. Since the values of all N^i are equal (defined as N_r , the product function representing the number of repeat units reduces to a simple exponential form. Therefore, the molecular formula may be expressed in simpler form as:

$$\left(\textcircled{I} (Z^c)_{N_c} \right) \left\{ \left(X^i Y^i (Z^i)_{N^i} \right)_{N_c N_r^{i-1}} \right\} \left(X^t Y^t (Z^t)_{N^t} \right)_{N_c N_r^{t-1}}$$

where $i = 1$ to $t-1$

This form still shows the distinction between the different generations i , which each consist of

$N_c N^{(i-1)}_r$ repeating units, $X^i Y^i (Z^i)_{N_i}$. Combining the generations into one term gives:

$$\left(\textcircled{I} (Z^c)_{N_c} \right) \left(X^r Y^r (Z^r)_{N_r} \right)_{N_c \frac{N_r^{(t-1)} - 1}{N_r - 1}} \left(X^t Y^t (Z^t)_{N_t} \right)_{N_c N_r^{(t-1)}}$$

or

$$\left(\textcircled{I} (Z^c)_{N_c} \right) \left\{ \left(X^r Y^r (Z^r)_{N_r} \right)_{\frac{N_r^{(t-1)} - 1}{N_r - 1}} \left(X^t Y^t (Z^t)_{N_t} \right)_{N_r^{(t-1)}} \right\}_{N_c}$$

Core Repeat Unit Terminal Unit

wherein $X^r Y^r (Z^r)_{N_r}$ is the repeating unit which is used
20 in all generations i.

Consequently, if a polymer compound will fit
into these above formulae, then the polymer is a
starburst polymer. Conversely, if a polymer compound
25 will not fit into these above formulae, then the
polymer is not a starburst polymer. Also, to determine
whether a polymer is a starburst polymer, it is not
necessary to know the process by which it was prepared,
but only whether it fits the formulae. The formulae
30 also demonstrate the generations (G) or tiering of
dendrimers.

Clearly, there are several ways to determine
the ratio of agent (M) to dendrimer (P) which depend
35 upon how and where the association of P*M occurs. When
there is interior encapsulation, the weight ratio of

M:P usually is 10:1, preferably 8:1, more preferably 5:1, most preferably 3:1. The ratio can be as low as 0.5:1 to 0.1:1. When interior stoichiometry is used, the weight ratio of M:P is the same as for interior
 5 encapsulation. When exterior stoichiometry is determined, the mole/mole ratio of M:P is given by the following formulae:

	M	:	P
10	<hr/>		
	(A) 5 $N_c N_t N_r^{G-1}$		1
	(B) 3 $N_c N_t N_r^{G-1}$		1
15	(C) 1 $N_c N_t N_r^{G-1}$		1

where N_c means the core multiplicity, N_t means the terminal group multiplicity, and N_r means branch
 juncture multiplicity. The $N_c N_t N_r^{G-1}$ term will result
 20 in the number of Z groups. Thus, for example, (A) above will result when proteins, enzymes or highly charged molecules are on the surface; (B) above when it is aspirin, 2,4-D or octanoic acid; (C) above when it
 25 is carboxylate ions or groups.

Of course other structures of various dimensions can be readily prepared by one skilled in the art by appropriately varying the dendrimer
 components and number of generations employed. A
 30 roughly scaled comparison of three different dendrimer series relative to an IgG antibody is seen in Figure 3. The series of drawings indicated by Figure 3(B) I shows the starburst polyamidoamines (PAMAM); by II shows the
 35 starburst polyethers (PE); and by III shows the starburst polyethyleneimines (PEI). In a manner

similar to that of Figure 1, all three series (I, II and III) have their far left drawing showing the initiator core, the next drawing from the left showing the starbranch oligomer, and the remaining drawings showing the starburst oligomers and respective starburst bridged dendrimers. It can be seen that in these series of scale drawings that the dendrimer dimensions are close to those noted for the IgG antibody Figure 3(A). The IgG antibody is shown to the far left in Figure 3. The scale is $1 \text{ mm} = 3.5 \text{ \AA}$. In Figure 3(A) the variable region is shown by (A); the constant region by (B); and the carbohydrate attachment sites by (C). Approximate measurements shown on Figure 3 are (1) is $35\text{--}40 \text{ \AA}$; (2) is 70 \AA ; and (3) is 60 \AA . These dimensional properties are preferred for instance where targeting involves exiting from the vascular system. Therefore dendrimer diameters of 125 Angstrom units or less are particularly preferred in that they may allow exiting from the vascular system to targeted organs serviced by continuous or fenestrated capillaries. These dimensions are significant in that they are small compared to the size of a potential targeting component such as an antibody (see Figure 3). A linear polymer of comparable molecular weight would have a radius of gyration, (in its fully extended form), that would be much larger than the same molecular weight dendrimer. A linear polymer of this type would be expected to adversely affect the molecular recognition properties of many accepted targeting components. It is also desirable that the conjugates be of minimum molecular volume so as not to discourage, extravasation, e.g., by coupling Fab, Fab'

or other appropriate antibody fragment to low molecular volume dendrimers.

5 Dendrimers are desirable for the delivery of radionuclides or strongly paramagnetic metal ions to tumor sites because of their ability to chelate a number of metal ions in a small volume of space. Coupling to antibodies or antibody fragments which are specific for tumors may deliver a number of metals per 10 antibody, with only a single modification to the antibody.

Linking target directors to dendrimers is another aspect of the present invention. In preferred 15 embodiments of the present invention, particularly where it is desired to use an antibody as a target director, a reactive functional group such as a carboxyl, sulfhydryl, reactive aldehyde, reactive olefinic derivative, isothiocyanato, isocyanato, amino, 20 reactive aryl halide, or reactive alkyl halide can conveniently be employed on the dendrimer. The reactive functional groups can be introduced to the dendrimer using known techniques, for example:

25 (1) Use of a heterofunctional initiator (as a starting material for synthesizing the dendrimer) which has incorporated into it functional groups of different reactivity. In such heterofunctional initiator at 30 least one of the functional groups will serve as an initiation site for dendrimer formation and at least one of the other functional groups will be available for linking to a target director but unable to initiate dendrimer synthesis. For example, use of protected

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aniline to allow further modification of NH_2 without reacting NH_2 .

5 The functional group which will be available for linking to a target director may be part of the initiator molecule in any one of three forms; namely:

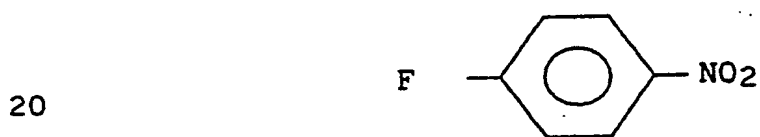
10 (a) In the form in which it will be used for linking with the target director. This is possible when none of the synthetic steps involved in the dendrimer synthesis can result in reaction at this center.

15 (b) When the functional group used for linking to the targeting director is reactive in the synthetic steps involved in the dendrimer synthesis, it can be protected by use of a protecting group, which renders the group unreactive to the
20 synthetic procedures involved, but can itself be readily removed in a manner which does not alter the integrity of the remainder of the macromolecule.

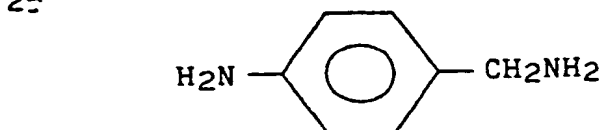
25 (c) In the event that no simple protecting group can be formed for the reactive functionality to be used for linking with the targeting director, a synthetic
30 precursor can be used which is unreactive in all the synthetic procedures used in the dendrimer synthesis. On completion of the synthesis, this functional group must be readily convertible into the desired
35 linking group in a manner which does not

alter the integrity of the remainder of the molecule.

(2) Coupling (covalently) the desired reactive functional group onto a preformed dendrimer, the reagent used must contain a functionality which is readily reacted with the terminal functional groups of the dendrimer. The functional group to be ultimately used to link with the targeting agent can be in its final form, as a protected functionality, or as a synthetic precursor. The form in which this linking functionality is used depends on its integrity during the synthetic procedure to be utilized, and the ability of the final macromolecule to withstand any conditions necessary to make this group available for linking. For example, the preferred route for PEI uses

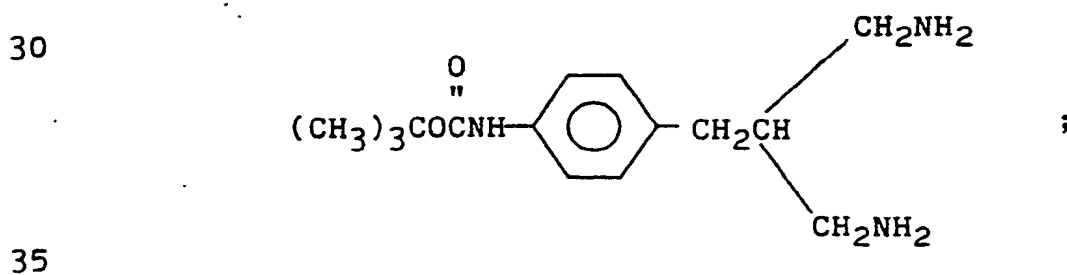
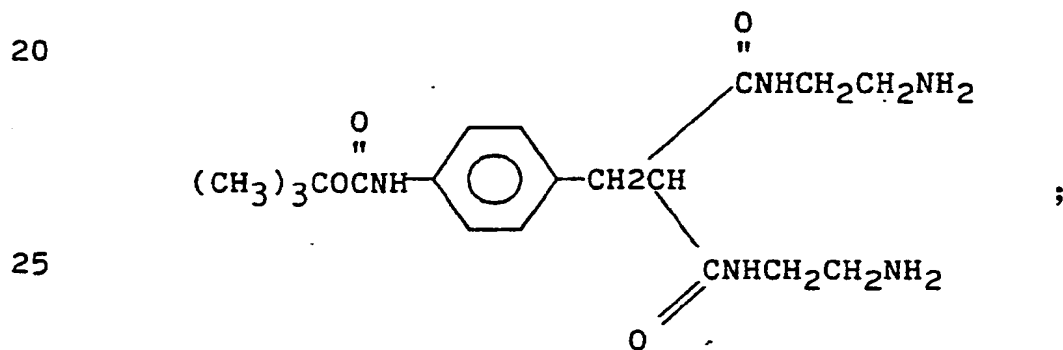
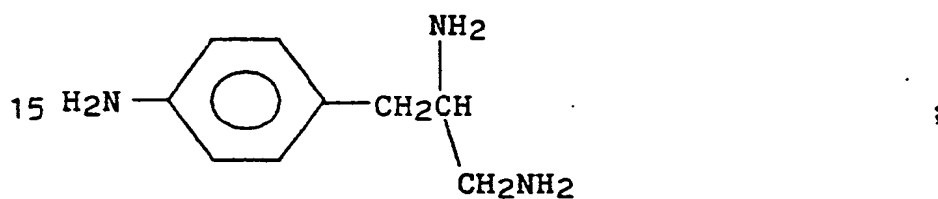
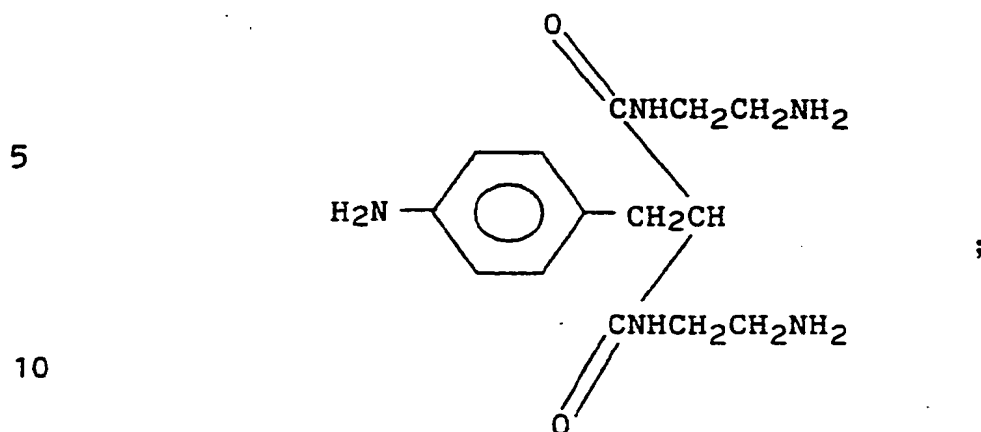


Examples of heterofunctional initiators for use in (1) above, include the following illustrative examples:

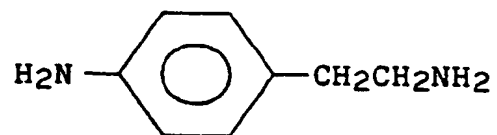


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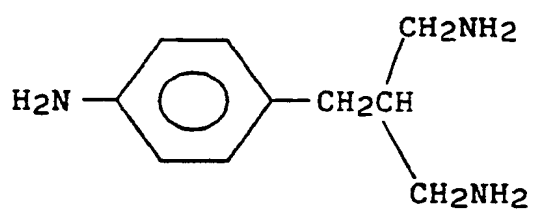


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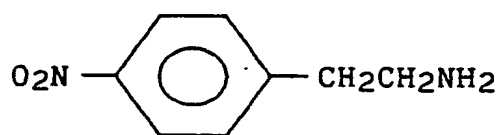
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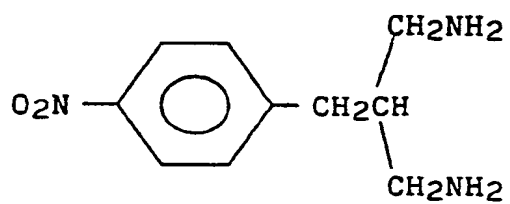
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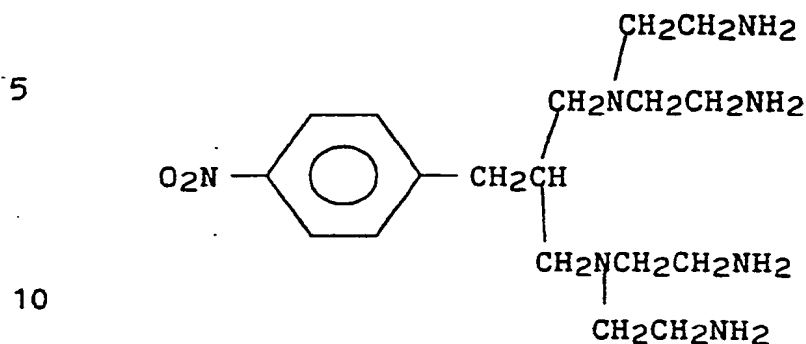
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; and

30

35



There are several chemistries of particular importance:

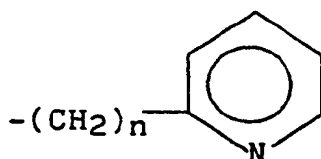
- 15
- 1) Starburst Polyamidoamides ("PAMAM") Chemistry;
 - 2) Starburst Polyethyleneimines ("PEI") Chemistry;
 - 3) Starburst PEI compound with a surface of PAMAM;
 - 20 4) Starburst polyether ("PE") chemistry.

Modifications of the dendrimer surface functionalities may provide other useful functional groups such as the following:

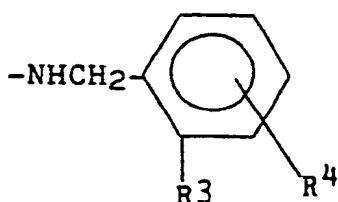
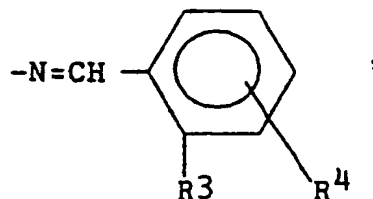
- 25 -OPO₃H₂, -PO₃H₂, -PO₃H⁽⁻¹⁾, -PO₃⁽⁻²⁾, -CO₂⁽⁻¹⁾, -SO₂H, -SO₂⁽⁻¹⁾, -SO₃H, -SO₃⁽⁻¹⁾, -NR¹R², -R⁵, -OH, -OR¹, -NH₂, polyethers, perfluorinated alkyl, -CNHR¹, -COH,
- O O

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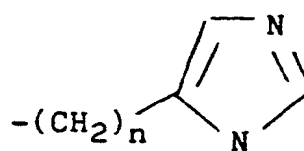
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5



10



wherein

15

R represents alkyl, aryl or hydrogen;

R¹ represents alkyl, aryl, hydrogen, or $-\text{N} \begin{array}{c} \text{(CH}_2\text{)}_n \\ \text{(CH}_2\text{)}_n \end{array} \text{X}$;

20

R² represents alkyl, aryl, or $-\text{N} \begin{array}{c} \text{(CH}_2\text{)}_n \\ \text{(CH}_2\text{)}_n \end{array} \text{X}$;

25

R³ represents -OH, -SH, -CO₂H, -SO₂H, or -SO₃H;

R⁴ represents alkyl, aryl, alkoxy, hydroxyl, mercapto, carboxyl, nitro, hydrogen, bromo, chloro, iodo, or fluoro;

30

R⁵ represents alkyl;

X represents NR, O or S; and

n represents the integer 1, 2 or 3;

35

polyethers; or other immuno insensitive moieties.

The choice of functional group depends upon the particular end use for which the dendrimer is designed.

5 Linking of antibodies to dendrimers is another aspect of the present invention. Typically, the antibodies or antibody fragments are linked to the dendrimer by techniques well known in the art such as attachment between a functional group on the dendrimer and moieties such as carbohydrate, amino, carboxyl, or
10 sulfhydryl functionalities present in the antibody or antibody fragment. In some instances connecting groups may be used as connectors or spacers between the dendrimer and antibody or antibody fragment. The
15 attachment of the dendrimer to the antibody or antibody fragment should be performed in a manner which does not significantly interfere with the immunoreactivity of the antibody or antibody fragment, that is, by binding the antibody or antibody fragment via a functionality
20 in the antibody or antibody fragment which is not a part of the antigen recognition and binding site.

 The following examples further illustrate the invention but are not to be construed as a limitation
25 on the scope of the invention. The lettered examples concern the preparation of starting materials; the numbered examples concern the preparation of products.

30

35

Example A: Preparation of 2-Carboxamido-3-(4'-nitro-phenyl)-propanamide.

p-Nitrobenzyl malonate diethylester (2.4 grams (g), 8.13 mmole) was dissolved in 35 ml of methanol. 5 The solution was heated to 50-55°C with stirring and a stream of anhydrous ammonia was bubbled through the solution for 64 hours. The solution was cooled and the white, flocculant product was filtered and 10 recrystallized from 225 milliliters (ml) of boiling methanol to afford 1.85 g (7.80 mmole) of bis amide in 96% yield (mp = 235.6°C(d)).

The structure was confirmed by MS, ¹H and ¹³C NMR spectroscopy. 15

Anal: Calc. for C₁₀H₁₁O₄N₃

	<u>C</u>	<u>H</u>	<u>N</u>
Theo:	50.63	4.69	17.72
20 Found:	50.75	4.81	17.94

Example B: Preparation of 1-Amino-2-(aminomethyl)-3-(4'-nitrophenyl)propane.

25 2-Carboxamido-3-(4'-nitrophenyl)propanamide (2.0 g, 8.43 mmole) was slurried in 35 ml of dry tetrahydrofuran under a nitrogen atmosphere with stirring. To this mixture was added borane/tetrahydrofuran complex (106 ml, 106 mmole) via syringe. The reaction mixture 30 was then heated to reflux for 48 hours during which time the suspended amide dissolved. The solution was cooled and the tetrahydrofuran was removed in vacuo using a rotary evaporator. The crude product and borane residue was dissolved in 50 ml of ethanol and 35 this solution was purged with anhydrous hydrogen chloride gas. The solution was refluxed for 1 hour and

the solvent removed in vacuo. The crude hydrochloride salt was dissolved in 15 ml of deionized water and extracted with two 50 ml portions of methylene chloride. The aqueous layer was cooled in an ice bath
5 under an argon blanket and 50% sodium hydroxide was slowly added until basic pH=11.7. The basic aqueous layer was extracted with four 25 ml portions of methylene chloride and these combined extracts were
10 evaporated (rotary) to give 1.45 g of amber colored oil. This oil was triturated with diethyl ether (50 ml) and filtered under pressure through a short silica gel (grade 62 Aldrich) column. The column was washed with 100 ml of ether and the combined filtrates were
15 vacuum evaporated giving 1.05 g (5.02 mmole) of the titled diamine as a clear oil (mp = 275-278°C(d) bis HCl salt).

The structure was confirmed by MS, ¹H and ¹³C NMR
20 spectroscopy.

Anal: Calc. for C₁₀H₁₇N₃O₂Cl₂

	<u>C</u>	<u>H</u>	<u>N</u>
25 Theo:	42.57	6.07	14.89
Found:	43.00	6.14	15.31

Example C: Preparation of 1-Amino-2-(aminomethyl)-3-(4'-aminophenyl)propane.

30 Borane/tetrahydrofuran solution (70 ml, 70 mmole) was added under nitrogen via a cannula needle to a flask containing 4-amino-benzyl malonamide (1.5 g, 7.24 mmole) with stirring. The solution was brought to
35 reflux for 40 hours. The colorless solution was cooled and excess tetrahydrofuran was removed by rotary evaporation leaving a clear gelatinous oil. Methanol

(50 ml) was cautiously added to the oil with notable gas evolution. Dry hydrogen chloride was bubbled through the suspension to effect dissolution and the solution was then refluxed for 1 minute. The methanol/HCl was rotary evaporated and the resulting hydrochloride salt was carried through the same dissolution/reflux procedure again. The hydrochloride salt obtained was dissolved in 10 ml of water and cooled in an ice bath under argon. Concentrated sodium hydroxide (50%) was added slowly with stirring to pH=11. The aqueous portion was then extracted with 2 X 100 ml portions of chloroform which were combined and filtered through a short silica gel plug without drying. The solvent was removed in vacuo (rotary) affording the title compound (0.90 g, 5.02 mmole) in 70% yield (R_f =0.65 - $\text{CHCl}_3/\text{MeOH}/\text{NH}_4\text{OH}$ conc - 2/2/1). The structure was confirmed by ^1H and ^{13}C NMR and used without further purification.

Example D: Preparation of 6-(4-Aminobenzyl)-1,4,8,11-tetraaza-5,7-dioxoundecane.

4-Aminobenzyl malonate dimethylester (2.03 g, 8.43 mmole) was dissolved in 10 ml of methanol. This solution was added dropwise to a stirred solution of freshly distilled ethylene diamine (6.00 g, 103.4 mmole) in 10 ml of methanol under nitrogen over a 2 hour period. The clear solution was stirred for 4 days and Thin Layer Chromatography (TLC) analysis indicated total conversion of diester (R_f = 0.91) to the bis amide (R_f = 0.42 - 20% conc $\text{NH}_4\text{OH}/80\%$ ethanol). This material was strongly ninhydrin positive. The methanol and excess diamine were removed on a rotary evaporator and the resulting white solid was vacuum dried (10^{-1} mm, 50°C) overnight to afford crude product (2.45g, 8.36

mmole) in 99% yield. An analytical sample was recrystallized from chloroform/hexane, MP = 160-161°C. The mass spectral, ¹H and ¹³C NMR data were consistent with the proposed structure.

5

Example E: Reaction of Mesyl Aziridine with 1-Amino-2-(aminomethyl)-3-(4-nitrophenyl)propane.

1-Amino-2-(aminomethyl)-3-(4-nitrophenyl)-
10 propane (400 mg, 1.91 mmole, >96% pure) was dissolved in 10.5 ml of absolute ethanol under nitrogen. Mesyl aziridine (950 mg, 7.85 mmole) was added to the stirred diamine solution as a solid. The reaction was stirred at 25°C for 14 hours using a magnetic stirrer and during
15 this period a white, gummy residue formed on the sides of the flask. The ethanol was decanted and the residue was triturated with another 15 ml portion of ethanol to remove any unreacted aziridine. The gummy product was vacuum dried (10¹mm, 25°C) to afford the tetrakis methyl
20 sulfonamide (1.0 g, 1.44 mmole) in 75% yield (R_f = 0.74 - NH₄OH/ethanol - 20/80). The structure was confirmed by ¹H and ¹³C nuclear magnetic resonance (NMR) spectroscopy.

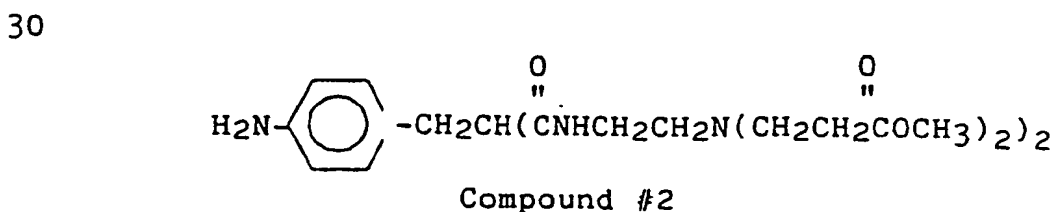
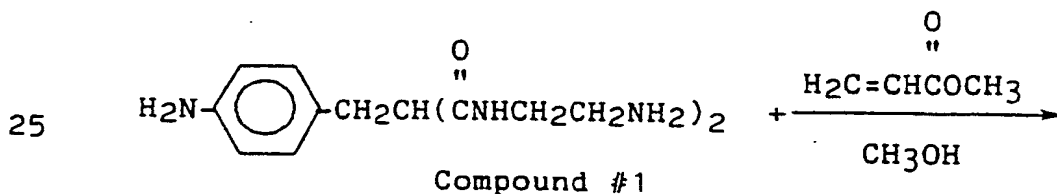
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Example F: Preparation of 2-(4-Nitrobenzyl)-1,3-(bis-N,N-2-aminoethyl)diaminopropane.

The crude methylsulfonamide (650 mg, 0.94
30 mmole) was dissolved in 5 ml of nitrogen purged, concentrated sulfuric acid (98%). This solution was maintained under nitrogen and heated to 143-146°C for 27 minutes with vigorous stirring. A slight darkening was noted and the cooled solution was poured into a stirred
35 solution of ether (60 ml). The precipitated white salt cake was filtered and immediately dissolved in 10 ml of

deionized water. The pH of the solution was adjusted to pH=11 with 50% NaOH under argon with cooling. The resulting solution was mixed with 90 ml of ethanol and the precipitated inorganic salts were filtered. The solvent was removed from the crude amine under reduced pressure and to the resulting light brown oil was added 190 ml of toluene under nitrogen. The mixture was stirred vigorously and water was removed through azeotropic distillation (Dean-Stark trap) until the remaining toluene acquired a light yellow color (30-40 ml remaining in pot). The toluene was cooled and decanted from the dark, intractable residues and salt. This solution was stripped of solvent in vacuo and the resulting light yellow oil was vacuum dried (0.2 mm, 35°C) overnight affording 210 mg of the product (60%) which was characterized by MS, ¹H and ¹³C NMR.

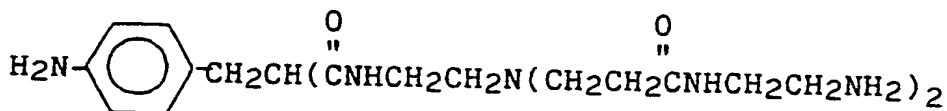
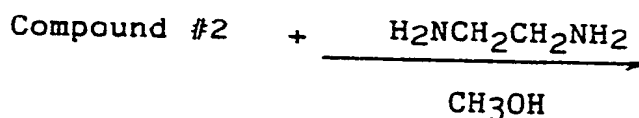
Example G: Preparation of a starburst polymer (containing an aniline derivative) of one half generation represented by the following scheme:



35 Methyl acrylate (2.09 g, 24 mmole) was dissolved in methanol (15 ml). The compound 6-(4-

aminobenzyl)-1,4,8,11-tetraaza-5,7-dioxoundecane (1.1 g, 3.8 mmole) (i.e., Compound #1) was dissolved in methanol (10 ml) and was added slowly over 2 hours with rigorous stirring to the methyl acrylate solution. The reaction mixture was stirred for 48 hours at ambient temperatures. The solvent was removed on the rotary evaporator maintaining the temperature below 40°C. The ester (Compound #2) was obtained as a yellow oil (2.6 g). No carboxyethylation of the aniline function was observed.

Example H: Preparation of a starburst polymer (containing an aniline moiety) of one generation; represented by the following scheme:

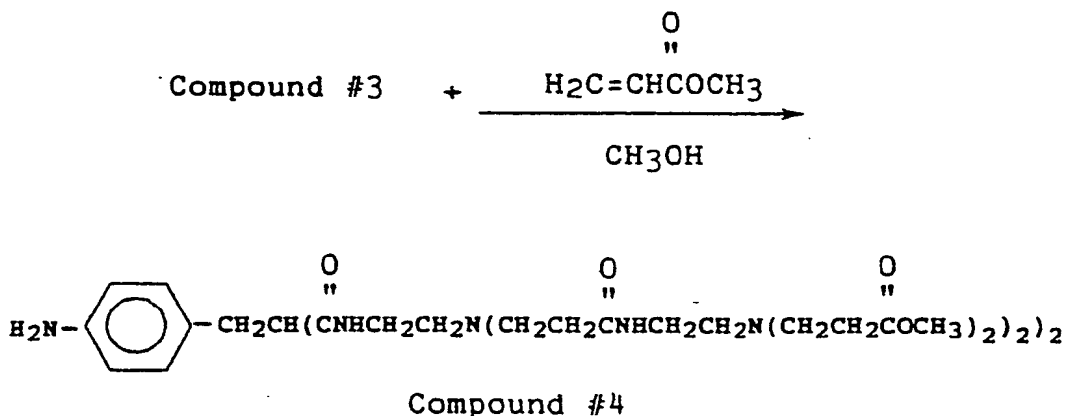


Compound #3

The ester (Compound #2) (2.6 g, 3.7 mmole) was dissolved in methanol (100 ml). this was carefully added to a vigorously stirring solution of ethylene diamine (250 g, 4.18 mole) and methanol (100 ml) at such a rate that the temperature did not rise above 40°C. After complete addition the reaction mixture was stirred for 28 hours at 35-40°C (heating mantle). After 28 hours no ester groups were detectable by infrared spectroscopy. The solvent was removed on the rotary evaporator at 60°C. The excess ethylene diamine was

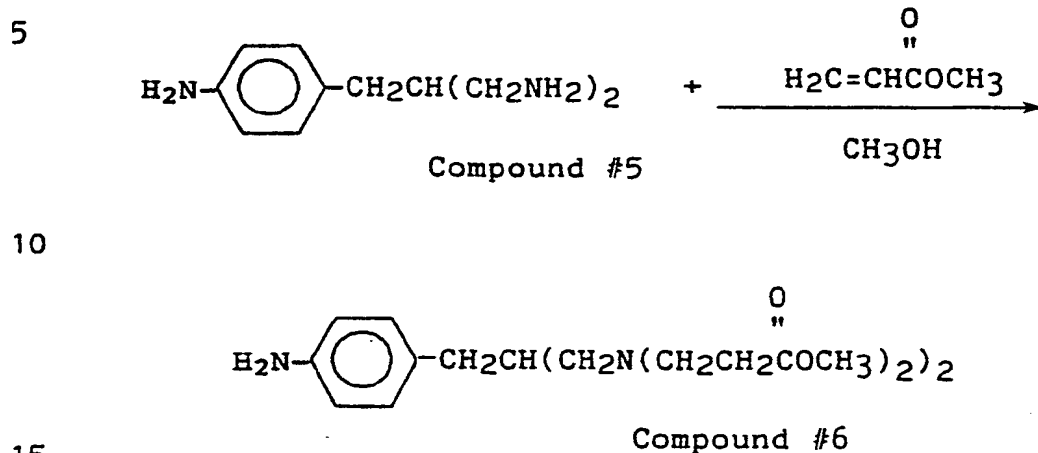
removed using a ternary azeotrope of toluene-methanol-ethylene diamine. Finally all remaining toluene was azeotroped with methanol. Removal of all the methanol yielded 3.01 g of the product (Compound #3) as an orange glassy solid.

Example I: Preparation of a starburst polymer (containing an aniline moiety) of one and one half generations represented by the following scheme:



The amine (Compound #3) (2.7 g, 3.6 mmole) was dissolved in methanol (7 ml) and was added slowly over one hour to a stirred solution of methyl acrylate (3.8 g, 44 mmole) in methanol (15 ml) at ambient temperatures. A slight warming of the solution was observed during the addition. The solution was allowed to stir at ambient temperatures for 16 hours. The solvent was removed on the rotary evaporator at 40°C. After removal of all the solvent and excess methyl acrylate the ester (Compound #4) was obtained in 4.7 g yield as an orange oil.

Example J: Preparation of a starburst polymer (containing an aniline moiety) of one half generation represented by the following scheme:

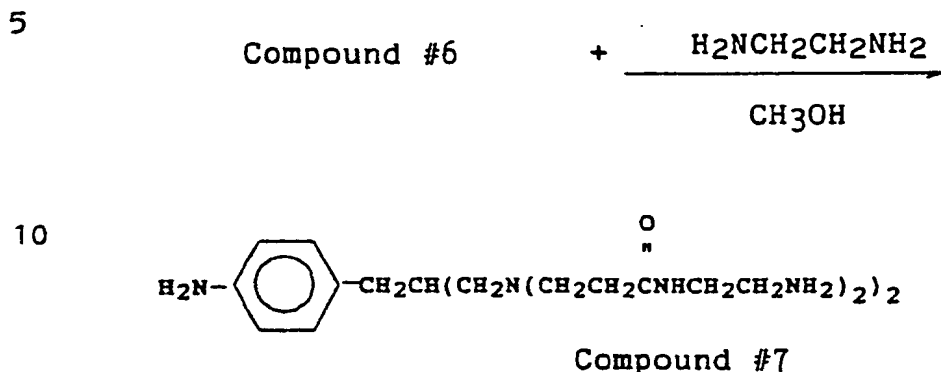


The triamine (Compound #5, the preparation of this compound is shown in Example C) (0.42 g, 2.3 mmole) was dissolved in methanol (10 ml) and was added dropwise over one hour to methyl acrylate (1.98 g, 23 mmole) in methanol (10 ml). The mixture was allowed to stir at ambient temperatures for 48 hours. The solvent was removed on the rotary evaporator, maintaining the temperature at no higher than 40°C. The excess methyl acrylate was removed by repeated azeotroping with methanol. The ester (Compound #6) was isolated as an orange oil (1.24 g).

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Example K: Preparation of a starburst polymer (containing an aniline moiety) of one generation; represented by the following scheme:



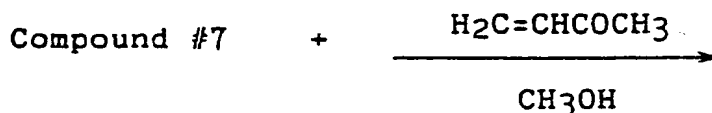
15 The ester (Compound #6) (1.24 g, 2.3 mmole) was dissolved in methanol (50 ml) and was added dropwise over two hours to ethylenediamine (73.4 g, 1.22 mole) in methanol (100 ml). A small exotherm was noted, 20 vigorous stirring was maintained. The solution was left to stir at ambient temperatures for 72 hours. The solvent was removed on the rotary evaporator at 60°C. The excess ethylene diamine was removed using a ternary azeotrope of toluene-methanol-ethylenediamine. Finally 25 all remaining toluene was removed with methanol and then pumping down with a vacuum pump for 48 hours gave the amine (Compound #7) (1.86 g) as a yellow/orange oil.

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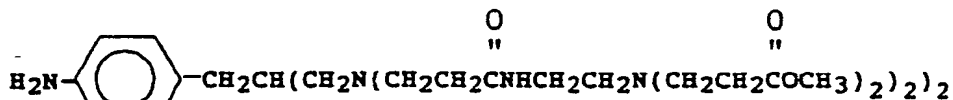
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Example L: Preparation of a starburst polymer (containing an aniline moiety) of one and one half generations; represent by the following scheme:

5



10



Compound #8

15

The amine (Compound #7) (1.45 g, trace of methanol remained) was dissolved in methanol (100 ml) and was added slowly over 1½ hours to a stirred solution of methyl acrylate (5.80 g) in methanol (20 ml). The solution was allowed to stir for 24 hours at room temperature. Removal of the solvent followed by repeated azeotroping with methanol enabled the removal of all the excess methyl acrylate. After pumping down on a vacuum pump for 48 hours the ester (Compound #8) was isolated as an orange oil (2.50 g, 1.8 mmole).

Example M: Hydrolysis of 4.5 generation dendrimer and preparation of calcium salt.

4.5 Generation PAMAM (ester terminated, initiated off NH₃) (2.11 g, 10.92 meq) was dissolved in 25 ml of methanol and to it was added 10% NaOH (4.37 ml, 10.92 meq) (pH = 11.5-12). After 24 hours at room temperature, the pH was about 9.5. After an additional

20 hours, the solution was rotovaped (rotary evaporated), 50 ml of toluene added, and rotovaped again.

5 The resulting oil was dissolved in 25 ml of methanol and precipitated as a white gum upon addition of 75 ml of diethyl ether. The liquid was decanted, and the gum was rotovaped to give a very fine off-white powder which upon further drying gives 2.16 g of
10 product (98% yield). No ester groups were found upon NMR and infrared analysis.

The sodium salt of 4.5 Generation PAMAM (ester terminated, initiated from NH_3) was replaced by the calcium salt via dialysis. The sodium salt (1.03 g) was dissolved in 100 ml of water and passed through hollow fiber dialysis tubing (cut off = 5000) at 3 ml/minute. The exterior of the tubing was bathed in 5% CaCl_2 solution. This procedure was then repeated.

The resulting solution was again dialyzed, this time against water, then repeated two additional times.

Evaporation provided 0.6 g of wet solid, which
25 was taken up in methanol (not totally soluble) and is
dried to give 0.45 g of off-white crystals.

30 C₃₆₉H₅₉₂O₁₄₁N₉₁Ca₂₄ Calc. - 10.10% Ca⁺⁺
M Wt. = 9526.3 Calc. = C-4432.1, H-601.8, O-2255.9,
N-1274.6, Ca-961.9)

Theo: C-46.5, H-6.32, N-13.38, Ca-10.10
Found: C-47.34, H-7.00, N-13.55, Ca-8.83

Example N: Preparation of dendrimers with terminal carboxylate groups.

Half-generation starburst polyamidoamines were hydrolyzed to convert their terminal methyl ester groups to carboxylates. This generated spheroidal molecules with negative charges dispersed on the periphery. The dendrimers hydrolyzed ranged from 0.5 generation (three carboxylates) to 6.5 generation (192 carboxylates).

10

The products could be generated as Na^+ , K^+ , Cs^+ or Rb^+ salts.

Example O: N-t-butoxycarbonyl-4-aminobenzyl malonate dimethylester

15

4-Aminobenzyl malonate dimethylester (11.62 g, 49 mmol) was dissolved in 50 ml of t-butanol:water (60:40 with stirring. Di-t-butoxydicarbonate (19.79 g, 90 mmol) was added and the reaction mixture stirred overnight. The butanol was removed on the rotary evaporator, resulting in a yellow suspension of the product in water. Extraction into methylene chloride, drying (MgSO_4) and evaporation gave a yellow oil (21.05 g, contaminated by di-t-butoxydicarbonate). Recrystallization from 2-propanol:water (75:25) yield pale yellow crystals (11.1 g, 33 mmol, 67%). The structure was confirmed by ^{13}C NMR and purity checked by HPLC analysis (Spherisorb ODS-1, 0.05M H_3PO_4 pH 3: CH_3CN 55:45). The material was used without further purification.

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Example P: N-t-butoxycarbonyl-6-(4-aminobenzyl)-
-1,4,8,11-tetraaza-5,7-dioxoundecane

N-t-butoxycarbonyl-4-aminobenzyl malonate dimethylester (8.82 g 26 mmol), prepared in Example O, was dissolved in 50 ml of methanol. This solution was added dropwise (2 hours) to a solution of freshly distilled ethylenediamine (188 g 3.13 mole) and 20 ml of methanol, under a nitrogen atmosphere. The solution was allowed to stir for 24 hours. The ethylene diamine/methanol solution was removed on the rotary evaporator. The product was dissolved in methanol and toluene added. Solvent removal on the rotary evaporator gave the crude product as a white solid (10.70 g contaminated with ethylenediamine). The sample was divided into two samples for purification. Azeotropic removal of ethylenediamine with toluene, using a soxhlet extractor with sulphonated ion exchange beads in the thimble to trap the ethylenediamine, resulted in partial decomposition of the product, giving a brown oil. The remaining product was isolated as a white solid from the toluene on cooling (2.3 g approximately 50 percent). Analysis of a 10 percent solution in methanol by gas chromatography (Column, Tenax 60/80) showed no ethylenediamine detectable in the sample (<0.1 percent). The second fraction was dissolved in methanol to give a 10 percent solution (by weight) and purified from the ethylenediamine by reverse osmosis, using methanol as the solvent. (The membrane used was a Filmtec FT-30, in an Amicon TC1R thin channel separator, the ethylenediamine crossing the membrane.) The product was isolated as a white solid (2.7 g), in which no detectable amounts of ethylenediamine could be found by gas chromatography. The ¹³C NMR data and HPLC analysis (Spherisorb ODS-1,

0.05M H₃PO₄ pH 3:CH₃CN 55:45) were consistent with the proposed structure. The product was used with no further purification.

- 5 Example Q: Preparation of a starburst dendrimer of one half generation from N-t-butoxycarbonyl-6-(4-aminobenzyl)-1,4,8,11-tetraaza-5,7-dioxoundecane

 N-t-butoxycarbonyl-6-(4-aminobenzyl)-1,4,8,11-
10 -tetraaza-5,7-dioxoundecane (5.0 g 13 mmol), prepared in Example P, was dissolved in 100 ml of methanol. Methyl acrylate (6.12 g, 68 mmol) was added and the solution stirred at ambient temperatures for 72 hours. The reaction was monitored by HPLC (Spherisorb ODS1, Acetonitrile: 0.04M Ammonium acetate 40:60) to optimize
15 conversion to the desired product. The solution was concentrated to 30 percent solids, and methyl acrylate (3.0 g 32 mmol) was added. The reaction mixture was stirred at ambient temperatures until no partially
20 alkylated products were detectable by HPLC (24 hours). Removal of the solvent at 30°C by rotary evaporation, and pumping down at 1 mm Hg for 24 hours gave the product as yellow viscous oil, yield 7.81 g. The ¹³C
25 NMR data was consistent with the proposed structure. The product was used without further purification.

- Example R: Preparation of a starburst dendrimer of one full generation from N-t-butoxycarbonyl-6-(4-aminobenzyl)-1,4,8,11-tetraaza-5,7-dioxoundecane

30 The half generation product (Example Q) (7.70 g, 10.45 mmol) was dissolved in 75 ml of methanol and added dropwise over 2 hours to a stirred solution of ethylenediamine (400 ml, 7.41 mol) and methanol (50
35 ml). The reaction mixture was stirred at ambient temperatures for 48 hours. The ethylenediamine and

methanol were removed by rotary evaporation to give a yellow oil (11.8 g contaminated with ethylenediamine). The product was dissolved in 90 ml of methanol, and purified from the ethylenediamine by reverse osmosis (Filmtec FT-30 membrane and Amicon TC1R thin channel separator, methanol as solvent). After 48 hours, no ethylenediamine could be detected by gas chromatography (Column, Tenax 60/80). Removal of the solvent on the rotary evaporator, followed by pumping down on a vacuum line for 24 hours gave the product as a yellow glassy solid (6.72 g). Analysis by HPLC, PLRP-S column, acetonitrile:0.015M NaOH, 10-20 percent gradient in 20 min.) and ^{13}C NMR analysis was consistent with the proposed structure.

Example S: Preparation of a starburst polymer of one and one half generation from N-t-butoxycarbonyl-6-(4-aminobenzyl)-1,4,8,11-tetraaza-5,7-dioxoundecane

The one generation product (Example R) (2.14 g, 25 mmol) was dissolved in 12.5 ml of methanol, and methyl acrylate (3.5 g, 39 mmol) in 5 ml of methanol was added. The solution was stirred at ambient temperatures for 48 hours, monitoring the progress of the reaction by HPLC (Spherisorb ODS-1, acetonitrile: 0.04M ammonium acetate, 60:40). A second aliquot of methyl acrylate was added (3.5 g 39 mmol) and the reaction mixture stirred at ambient temperatures for a further 72 hours. Removal of the solvent on the rotary evaporator gave the product as a yellow oil (3.9 g) after pumping down overnight with a vacuum pump. The product was used with no further purification.

Example T: Preparation of a starburst polymer of two full generations from N-t-butoxycarbonyl-6-(4-aminobenzyl)-1,4,8,11-tetraaza-5,7-dioxoundecane

The one and one half generation product (Example S) (3.9 g, 2.5 mmol) was dissolved in 50 ml of methanol, and was added dropwise over 2 hours to a stirred solution of ethylenediamine (600 g, 10 mol) and methanol (50 ml). The solution was stirred at ambient temperatures under an atmosphere of nitrogen for 96 hours. The ethylenediamine/methanol was removed on the rotary evaporator to give a yellow glassy solid (4.4 g contaminated with ethylenediamine). A 10 percent solution of the product was made in methanol, and purified from the ethylenediamine by reverse osmosis (membrane used as a Filmtex FT-30, in an Amicon TC1R thin channel separator) until no ethylenediamine could be detected by gas chromatography (Column, Tenax 60/80. Removal of the solvent gave the product as a yellow glassy solid (3.52 g). The ^{13}C NMR data and HPLC analysis (PLRP-S column, acetonitrile:0.015 M NaOH, 10 to 20 percent gradient in 20 minutes, were consistent with the proposed structure.

Example U: Reaction of the two generation starburst with Bromoacetic Acid to give a methylene carboxylate terminated starburst dendrimer

The second generation product (Example T) (0.22 g, 0.13 mmol) was dissolved in 15 ml of deionized water and the temperature equilibrated at 40.5°C. Bromoacetic acid (0.48 g, 3.5 mmol) and lithium hydroxide (0.13 g, 3.3 mmol) were dissolved in 5 ml of deionized water, and added to the reaction mixture. The reaction pH was carefully maintained at 9, with the use of a pH stat (titrating with 0.1N NaOH), at 40.5°C overnight.

Monitoring by reverse phase HPLC, (Spherisorb ODS-1 column, eluent 0.25 M H_3PO_4 pH 3 [NaOH]; acetonitrile 85:15) confirmed the synthesis of predominantly a single component.

5

Example V: Preparation of Isothiocyanato functionalized second generation methylene-carboxylate terminated starburst dendrimer

10 Five ml of a 2.8 mM solution of the second generation methylenecarboxylate terminated starburst dendrimer (Example U) was diluted with 20 ml water and the pH adjusted to 0.5 with concentrated hydrochloric acid. After one hour at room temperature the mixture
15 was analyzed by HPLC to verify the removal of the butoxycarbonyl group and then treated with 50 percent sodium hydroxide to bring the pH to 7. A pH stat (titrating with 0.1 N NaOH) was used to maintain the pH at 7 and 225 μl thiophosgene was added. After 15
20 minutes at room temperature the pH of the mixture was adjusted to 5 with 1N HCl. The mixture washed with chloroform (20 ml x 2) then concentrated on a rotary evaporator at reduced pressure. The residue recovered
25 0.91 g is a mixture of the isothiocyanate and salts.

Example W: Preparation of second generation starburst polyethyleneimine-methane sulfonamide

30 To a solution of 125 g N-methanesulfonyl-aziridine in 50 ml ethanol was added 25.0 g tris(2-aminoethyl)amine. The solution was stirred at room temperature for 4 days. Water was added to the reaction mixture as needed to maintain the homogeneity of the solution. The solvent was removed by
35 distillation in vacuo to give the 2nd generation

starburst PEI-methane sulfonamide as a yellow glass (161 g).

5 Example X: Cleavage of methane sulfonamides to form second generation starburst polyethyleneimine

 A solution of 5.0 g of second generation starburst PEI-methane sulfonamide, from Example W in 20 ml of 38 percent HCL was sealed in a glass ampoule. 10 The ampoule was heated at 160°C for 16 hours, then cooled in an ice bath and opened. The solvent was removed by distillation in vacuo and the residue dissolved in water. After adjusting the pH of the solution to greater than or equal to 10 with 50 percent 15 NaOH, the solvent was removed by distillation in vacuo. Toluene (150 ml) was added to the residue and the mixture heated at reflux under a Dean-Stark trap until no more water could be removed. The solution was filtered to remove salts and the filtrate concentrated 20 in vacuo to give 1.9 g second generation starburst PEI as a yellow oil.

25 Example Y Preparation of third generation starburst polyethyleneimine-methane sulfonamide

 To a solution of 10.1 g second generation starburst PEI, from Example X, in 100 ml ethanol was added 36.6 g N-methanesulfonylaziridine. The solution was stirred at room temperature for 1 week. Water was 30 added as needed to maintain the homogeneity of the solution. The solvent was removed by distillation in vacuo to give third generation starburst PEI-methane sulfonamide as a yellow glass (45.3 g).

35

Example Z: Cleavage of methane sulfonamides to form 3rd generation starburst polyethyleneimine

The methane sulfonamide groups of third generation starburst PEI-methane sulfonamide (5.0 g),
5 from Example Y, were removed by the same procedure as described for the second generation material in Example X to give 2.3 g third generation starburst PEI as a yellow oil.

10 Example AA: Reaction of a third generation starburst polyethyleneimine with 4-fluoro-nitrobenzene

The third generation starburst polyethyleneimine (Example Z) (1.06 g, 1.2 mmol) was
15 dissolved in 12 ml of absolute ethanol. (4-Fluoro)-nitrobenzene (120 μ l, 1.2 mmol) was added and the reaction mixture refluxed overnight. The solvent was removed on the rotary evaporator, and the bright yellow oil dissolved in water. The aqueous solution was
20 washed with chloroform to remove any unreacted (4-fluoro)-nitrobenzene. Removal of the water gave the product as a deep yellow oil (0.80 g). The ^{13}C NMR spectrum was consistent with the proposed structure.
25 (No attempt was made to determine the nature of the statistical distribution). The product was used without further purification.

30 Example BB: Reaction of the nitrophenyl derivative of the third generation starburst polyethyleneimine with glycolonitrile.

The nitrophenyl derivative of the third generation starburst polyethyleneimine (Example AA) (0.80 g) was dissolved in 20 ml of deionized water.
35 Sodium hydroxide (2.80 g, 50 percent w/w) was added to the stirred solution, and the solution purged with

nitrogen, venting through a sodium hydroxide scrubber. Glycolonitrile (2.85 ml of a 70 percent aqueous solution) was added at ambient temperatures. A yellow precipitate was observed to form after a few minutes.

5 After two hours, the temperature was slowly raised to a reflux, and the solution maintained at a reflux with a nitrogen purge for 24 hours. Removal of the water gave the product as a yellow solid contaminated with glycolic acid and sodium hydroxide. The ^{13}C NMR

10 spectrum was consistent with the proposed structure. The product was used without further purification.

Example CC: Hydrogenation of the nitrophenyl derivative to the aminophenyl methylenecarboxylate terminated third generation starburst polyethyleneimine.

15

The yellow solid from Example BB (1.70 g) was dissolved in 10 ml of deionized water, the resulting pH of the solution was approximately 11. Palladium on charcoal (200 mg of 5 percent Pd/C) was added to the

20 reaction mixture in a glass Parr shaker bottle. The reaction mixture was placed under a pressure of 40 psi (275 kPa) of hydrogen, and shaken at ambient temperature in a Parr hydrogenation apparatus, for 6

25 hours. The reaction mixture was then filtered through a 0.5 m Millipore filter to remove the Pd/C and the solvent removed in vacuo and was gel filtered through a Biogel P2 resin (25 g swollen with water).

30 Acidification with HCl resulted in an orange brown solution, which was purged with nitrogen overnight. Removal of the solvent in vacuo gave the product as the hydrochloride salt which was a pale brown solid (3.98 g, contaminated with NaCl and glycolic acid, maximum

35

theoretical amount of product 1.15g). The product was used with no further purificate.

5 Example DD: Preparation of 4-isothiocyanatophenyl methylenecarboxylate terminated third generation starburst polyethyleneimine

10 The product from Example CC (3.98 g) was dissolved in 15 ml of deionized water and an aliquot (2.5 ml) of this solution was diluted with 10 ml water. the pH of the solution was adjusted to 7 with sodium hydroxide. A pH stat (titrating with 1N NaOH) was used to maintain the pH and 200 μ l thiophosgene was added. After 10 minutes the pH of the mixture was adjusted to 15 4 with hydrochloric acid. Water was removed on a rotary evaporator at reduced pressure (a small amount of n-butanol was added to prevent foaming) . The residue was washed with methylene chloride and then dried. The crude product (0.95 g) a mixture of 20 isothiocyanate (0.14 g) and salts was used without further purification.

25 Example EE: Preparation of a methylenecarboxylate-terminated second generation starburst polyamidoamine (initiated from ammonia)

30 The second generation starburst polyamidoamine (2.71 g, 2.6 mmol) and bromoacetic acid (4.39 g, 31.6 mmol) were dissolved in 30 ml of deionized water and the pH adjusted to 9.7 with 5N NaOH using a pH stat. The reaction was maintained at this pH for a half hour, and the temperature was slowly raised to 60°C and was maintained at 60°C for three hours at constant pH. The pH was raised to 10.3, and the reaction mixture 35 remained under control of the pH stat at ambient temperatures overnight. The reaction mixture was

refluxed for a further four hours prior to work up. Removal of the solvent, and azeotropeing the final traces of water with methanol gave the product as a pale yellow powder (8.7 g, contaminated with sodium bromide). The ^{13}C /NMR spectrum was consistent with the propose structure (with some contamination due to a small amount of defected material as a result of some monoalkylation).

10 Example FF: Preparation of a methylenecarboxylate terminated second generation starburst polyethyleneimine (initiated from ammonia)

15 The second generation starburst polyethyleneimine (2.73 g, 6.7 mmol), from Example X, and bromoacetic acid (11.29 g 81 mmol) were dissolved in 30 ml of deionized water. The pH was slowly raised to pH 9.5 maintaining the temperature below 30°C. The temperature was raised slowly to 55°C, and the reaction pH
20 maintained at 9.5 for 6 hours with the aid of a pH stat (titrating with 5N NaOH). The pH was raised to 10.2, and maintained at that pH overnight. Removal of the solvent on the rotary evaporator, and azeotropeing the final traces of water using methanol, gave the product
25 as a yellow powder (17.9 g, contaminated with sodium bromide). The ^{13}C NMR spectrum was consistent with the proposed structure (with some contamination due to a small amount of defected material as a result of some
30 monoalkylation).

Example GG: Preparation of 3.5, 4.5, 5.5 and 6.5 generation starburst PAMAM:

35 To a 10 weight percent solution of 2.46 g 3 generation PAMAM starburst was added 2.32 g of methyl acrylate. This mixture was allowed to sit at room

temperature for 64 hours. After solvent and excess methyl acrylate removal, 4.82 g of product was recovered (105% of theoretical).

5 Preparation of higher $\frac{1}{2}$ generation starburst
PAMAM's:

Generations 4.5, 5.5 and 6.5 were prepared as described above with no significant differences in
10 reactant concentrations, reactant ratios or reaction times.

Example HH: Preparation of 4, 5 and 6 generation starburst PAMAM:

15 To 2000 g of predistilled ethylenediamine was added 5.4 g of 4.5 generation starburst PAMAM as a 15 weight percent solution in methanol. This was allowed to sit at room temperature for 48 hours. The methanol and most of the excess ethylene diamine were removed by
20 rotary evaporation under water aspirator vacuum at temperature less than 60°C. The total weight of product recovered was 8.07 g. Gas chromatography indicated that the product still contained 34 weight percent
25 ethylenediamine at this point. A 5.94 g portion of this product was dissolved in 100 ml methanol and ultrafiltered to remove the residual ethylenediamine. The filtration was run using an Amicon TC1R thin channel recirculating separator equipped with an Amicon
30 YM2 membrane. An in-line pressure relief valve was used to maintain 55 psig (380 kPa) pressure across the membrane. The 100 ml was first concentrated to 15 ml by forcing solvent flow exclusively through the
35 membrane. After this initial concentration, the flow was converted to a constant volume retentate recycle

-75-

mode for 18 hours. After this time, 60 ml of methanol was passed over the membrane to recover product still in the module and associated tubing. The product was stripped of solvent and 2.53 g of 5 generation starburst PAMAM was recovered. Analysis by gas chromatography indicated 0.3 percent residual ethylenediamine remained in the product.

Preparation of generation 4 and 6 proceeded as above with the only difference being the weight ratio of ethylenediamine to starting material. To prepare 4th generation this ratio was 200:1 and for 6th generation this ratio was 730:1.

Example 1: Incorporation of 2-(acetyloxy)benzoic acid (aspirin) into starburst dendrimers.

A widely accepted method for ascertaining whether a "probe molecule" is included in the interior of a micelle is to compare its carbon-13-spin lattice relaxation times (T_1) in a non-micellized versus micellized medium. A substantial decrease in T_1 for the micellized medium is indicative of "probe molecule" inclusion in the micelle. Since starburst dendrimers are "covalently fixed" analogs of micelles, this T_1 relaxation time technique was used to ascertain the degree/extent to which various pharmaceutical type molecules were associated with starburst polyamidoamines. In the following examples, T_1 values for (acetyloxy)benzoic acid (I) (aspirin) were determined in solvent (CDCl_3) and then compared to T_1 values in CDCl_3 at various [I:dendrimer] molar ratios.

Inclusion of aspirin (I) into various starburst polyamidoamine dendrimers as a function of generation.

Various half generation (ester terminated, initiated from NH_3) starburst polyamidoamine dendrimers ($G = 0.5 \rightarrow 5.5$) were combined with 2-(acetyloxy)benzoic acid in CDCl_3 to give acid:tertiary amine ratios of = 1.0. A plot of T_1 values for 2-(acetyloxy)benzoic acid versus generation of starburst dendrimer added (see Figure 4 where \cdot represent C-4, π represent C-6, and o represent C-5) shows that T_1 reaches a minimum over the generation range of $2.5 \rightarrow 5.5$ for carbons 4, 5 and 6 in 2-(acetyloxy)benzoic acid. This demonstrates association of 2-(acetyloxy)benzoic acid in the dendrimers ($G = 2.5 \rightarrow 5.5$) and further confirms that

polyamidoamine dendrimers (Gen = 2.5 or greater) can function as carrier molecules.

5 Example 2 Release of pseudoephedrine from starburst dendrimer -PAMAM

 Pseudoephedrine (0.83 mg/ml) and starburst PAMAM dendrimer [1.0 mg/ml; G= 6.5; terminal group (Z) = 192 (methyl ester)] were dissolved in deionized
10 distilled water and the pH of the donor phase was adjusted to 9.5, with sodium hydroxide solution, and stored at room temperature for about 12 hours.
 Solution of pseudoephedrine alone was treated in the same way (control). The drug dendrimer solution was
15 stored at 40°C for 8 hours after the first experiment and dynamic dialysis performed. Dialysis membrane used was a SpectraPor 7, MWCO 1,000 28.6 mm in diameter in spectrum separation cells (half cell volume 5 and 10
20 ml, cell dimensions: 38 mm diameter for both the cells and the cell depth of 10 and 20 mm for 5 and 10 ml cells, respectively).

 Samples were analyzed by a HPLC procedure
25 developed for pseudoephedrine conditions for which are as follows:

 Column: uBondapak C-18
30 Mobile phase: pH 3.2 phosphate buffer plus acetonitrile (80:20)
 Flow rate : 0.3 ml/min
 Detection: UV at 210 nm
 Retention time: 13.3 min

35

The dialysis membrane was washed with deionized water and was kept soaking in the receptor phase for at least 12 hours prior to use. The dialysis membrane was placed in between the donor and the receptor compartment was stirred with a small magnetic spin bar. Known volumes of donor and receptor solutions were introduced into the respective compartments and transfer of pseudoephedrine to the receptor compartment was followed as a function of time. To maintain sink conditions the entire receptor phase was removed periodically (every 30 minutes) and replaced with fresh receptor phase. The amount of pseudoephedrine was assayed in the sampled receptor phase. Experiments were conducted at room temperature (22°C). The receptor phase was plain deionized distilled water.

The results of dynamic analysis are shown in Figure 5. In Figure 5, the • represents pseudoephedrine only (control), the (I) represents pseudoephedrine plus the dendrimer, and the O represents pseudoephedrine plus the dendrimer at 40°C, 8 hours before dialysis. It is apparent that in presence of G 6.5 dendrimer in the donor compartment the rate of dialysis of pseudoephedrine is reduced. Storing the donor solution at 40°C, appears to further reduce the rate of dialysis.

The experiment was repeated at lower with the concentrations (the ratio of number of drug molecules to the number of terminal groups was kept the same). G 6.5 dendrimer 120 µ/ml pseudoephedrine 100 µ/ml (122 µ/ml salt).

Dynamic dialysis of pseudoephedrine (alone) at this lower concentration was almost identical to that

at higher concentration. Figure 6 summarizes the results of this experiment where • represents pseudoephedrine only (control), and o represents pseudoephedrine plus dendrimer.

5

Example 3

The procedure of Example 2 was repeated using the modifications given below.

10

Receptor phase: pH 7.4 phosphate buffer
Donor phase: pH 7.4 phosphate buffer plus drug and dendrimer in the following ratios:

15

1. G 6.5 : Drug :: 1 : 192
2. G 5.5 : Drug :: 1 : 96
3. G 4.5 : Drug :: 1 : 48
4. G 6.5H: Drug :: 1 : 192
5. G 5.5H: Drug :: 1 : 96
6. G 4.5H: Drug :: 1 : 48

20

The above donor phase compositions plus pseudoephedrine alone were subjected to dynamic dialysis. The letter "H" after the dendrimer generation number stands for hydrolyzed dendrimer. Hydrolysis was accomplished by the procedure described in Examples M and N.

30

The results of these experiments are summarized in Figure 7 where the donor and receptor compartment contained pH 7.4 phosphate buffer. For pseudoephedrine alone (P) the mean curve of three experiments is plotted (shown by the solid line), and one typical run

35

from the other experiments are shown. In Figure 7, the following symbols represent the dendrimer of the indicated generation.

5		Table III
	Symbol	Dendrimer Generation
	●	5.5
10	●	6.5
	⊙	4.5
	⊖	5.5H
	○	6.5H
15	⊙	4.5H

Pseudoephedrine appears not to associate with the dendrimer (unhydrolyzed) at pH 7.4. Hydrolysis of the terminal functional groups into carboxylate form, has a dramatic effect on the dialysis rate (reduction). The generation number appears not to influence the dialysis rate.

25 Example 4: Interaction studies of salicylic acid with PAMAM starburst dendrimers

This example evaluated interaction characteristics of salicylic acid with PAMAM starburst dendrimers. These dendrimers consisted of an ammonia initiated core with repeating units derived from N-(2-aminoethyl) acrylamide. Both full (amine terminated functions) and half (ester terminal groups) generation polymers were included in the studies. The ratio of salicylic acid to starburst dendrimers utilized in the experiments resulted in approximately one salicylic

acid molecule to one terminal amine functional group for full generation polymers. In the half-generation polymer study, the same ratio was employed with adjustments made for the higher molecular weight polymer.

The experiments were conducted at room temperature using an equilibrium static cell dialysis methodology. Spectrum static dialysis cells (half cell volume, 10 ml) separated by SpectraPor 6 membranes (molecular weight cutoff = 1000) were utilized for all experiments. Transport of salicyclic acid was monitored as a function of time by removing aliquots from appropriate cell compartments and assayed by HPLC analysis using a U.V. detector at 296 nm (Bondupak C-18 Column, eluting mobile phase of acetonitrile/0.1M phosphate buffer (pH 3.2) at a ratio of 20:80 (V/V), set at a flow rate of 30 ml/hour).

Ten ml of a solution containing 1 mg/ml salicyclic acid and 2.5 mg/ml starburst polymer (Gen 4.0) adjusted to pH 6.65 and 5.0 with HCL solution were placed in the donor compartment of the dialysis cell and an equal volume of purified water adjusted to the same pH's placed in the receptor compartment. Transport of salicyclic acid into the receptor compartment was monitored. The results are given in Figure 8. In Figure 8, the free acid is represented by ●, the acid plus generation 4.0 dendrimer, pH 6.65 is represented by ○, and the acid plus generation 4.0 dendrimer, pH 5.00 is represented by □.

Due to the lower percent ionization of the amine groups on the polymer at pH 6, a greater extent of interaction with salicylic may be expected at pH 5,

resulting in less compound transported at the lower pH. The results given in Figure 8 indicate a much lower percentage of salicylic acid transported in the presence of polymer at both pH's studied compared to the salicylic acid control study. It is also observed that more salicylic acid is transported at pH 6.65 than at pH 5.0 as predicted. The data demonstrates an interaction of the starburst polymer with salicylic acid that can be controlled by pH. Sustained release characteristics are also implied by the data since the salicylic acid levels in the presence of polymer continue to rise past the approximate 12-hour equilibrium point observed in the control study.

To further investigate the interaction characteristics of salicylic acid with starburst polymers (Gen = 4.0) an experiment was designed at pH 8.0. The design of the study differed from that previously described in that only the salicylic acid solution (1 mg/ml), adjusted to pH 8.0, was placed in the donor compartment and the polymer solution (2.5 mg/ml) placed in the receptor compartment. Loss of salicylic acid from the donor compartment was monitored as previously described. The results of the experiment are given in Figure 9. In Fig 9, the free acid is represented by $-\bullet-$, and the acid plus generation 4.0 dendrimer at pH 8.0 is represented by $---\Delta---$.

As indicated in Figure 9, the equilibrium characteristics of salicylic acid in the donor compartment with starburst polymers in the receptor compartment differs from the salicylic acid control study. Based on the ionization characteristics of the molecules at pH 8, approximately 6-7% interaction is

expected. The observed extent of interaction is indicated to be on the order of 4-5%. The lower association observed may be due to experimental variability or to an ionization constant of less than one.

This experiment indicates an uptake or removal of free salicylic acid from the continuous phase of the system by the polymer. This type of action could result in suppression of reactivity of molecules suggesting a possible chelating type of property associated with the polymers.

The interaction characteristics of salicylic acid at pH 6.65 with a half generation starburst polymer (Gen = 4.5) having ester terminated functional groups were evaluated. Salicylic acid (1 mg/ml) was combined with starburst polymer (Gen = 4.5) 3.6 mg/ml at pH 6.65. Ten ml of the solution was placed in the donor compartment and transport from the donor compartment was monitored as previously described. The results are given in Figure 10. In Figure 10, the free acid is represented by -●-, and the acid plus polymer is represented by ---o---.

Under these experimental conditions no charge interaction is predicted to occur since the tertiary amine groups are non-ionized at pH 6.65. As is indicated in Figure 10, the loss of salicylic acid in the presence of polymer (Gen 4.5) is virtually identical during the first 10 hours of dialysis to that of the salicylic acid control study.

The following observations are made from the data presented in this example:

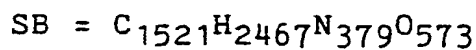
- (1) Full generation PAMAM starburst polymers function as a carrier for salicylic acid.
- 5 (2) Full generation PAMAM starburst polymers have sustained release functionality for salicylic acid.
- 10 (3) Salicylic acid carrier properties of full generation PAMAM starburst polymers can be controlled by pH.

Example 5: Demonstration of multiple chelation of iron by a sodium propionate terminated sixth generation starburst polyamidoamine.

15 The sodium propionate terminated sixth generation polyamidoamine (initiated from ammonia) (97.1 mg, 2.45 mol.) was dissolved in 1.5 ml of deionized water. Addition of 0.5 ml of 0.5N HCl reduced the pH to 6.3. Ferric chloride was added (0.5
20 ml of 0.1.2M solution, 0.051 mmol) producing a light brown gelatinous precipitate. On heating at 60°C for 0.5 hours, the gelatinous precipitate became soluble, resulting in a homogeneous orange solution. The
25 solution was filtered through Biogel P2 acrylamide gel (10 g, twice) isolating the orange band (free of halide contamination). Removal of the solvent in vacuo gave the product as an orange film (30 mg). Analysis was
30 consistent with chelation of approximately 20 moles of ferric ions per mole of starburst dendrimer.

Table IV

	Found	Theoretical		
		$\text{Na}_4\text{Fe}_{20}\text{H}_{128}\text{SB}$	$\text{Na}_5\text{Fe}_{20}\text{H}_{127}\text{SB}$	$\text{Na}_6\text{Fe}_{20}\text{H}_{126}\text{SB}$
Na	0.39, 0.24 (0.31 0.1%)	0.25	0.31	0.38
Fe	3.14, 3.11 (3.12 0.02%)	3.05	3.05	3.04
C	47.11	49.87	49.84	49.81
H	7.33	7.31	7.30	7.29
N	14.81	14.49	14.48	14.47
O	----	25.03	25.02	25.01
	Mwt.	36632.23	36654.21	36375.18



20 These results confirm chelation of 20 ± 2 moles of ferric ions per mole of starburst dendrimer.

Example 6: Preparation of a product containing more than one rhodium atom per starburst polymer.

25 2.5 Gen PAMAM (ester terminated, initiated from NH_3) (0.18 g, 0.087 mmole) and $\text{RhCl}_3 \cdot 3\text{H}_2\text{O}$ (0.09 g, 0.3 mmole) were mixed in dimethylformamide (DMF) (15 ml) and heated for 4 hours at 70°C . The solution turned crimson and most of the rhodium was taken up. The
30 unreacted rhodium was removed by filtration and the solvent removed on the rotary evaporator. The oil formed was chloroform soluble. This was washed with water and dried (MgSO_4) before removal of solvent to yield a red oil (0.18 g). The NMR spectrum was
35 recorded in CDCl_3 only minor differences were noted between the chelated and unchelated starburst.

Dilution of some of this CDCl_3 solution with ethanol followed by NaBH_4 addition resulted in rhodium precipitation. $\text{RhCl}_3 \cdot 3\text{H}_2\text{O}$ is insoluble in chloroform and in chloroform starburst solution thus confirming
5 chelation.

Example 7: Preparation of a product containing Pd chelated to a starburst polymer

10 3.5 Generation PAMAM (ester terminated, initiated from NH_3) (1.1 g, 0.24 mmole) was dissolved with stirring into acetonitrile (50 ml). Palladium chloride (0.24 g, 1.4 mmole) was added and the solution was heated at $70-75^\circ\text{C}$ (water bath) overnight. The PdCl_2
15 was taken up into the starburst. After removal of the solvent, the NMR in CDCl_3 confirmed that chelation had occurred. Dilution of the CDCl_3 solution with ethanol and addition of NaBH_4 resulted in precipitation of the palladium. The chelated product (1.23 g) was isolated
20 as a brown oil.

Example 8: Demonstration of multiple chelation of yttrium by a methylene carboxylate terminated second generation starburst polyethyleneimine by trans
25 chelation from yttrium acetate

The starburst polyethyleneimine methylene carboxylate terminated material (0.46 g 52.5 percent active, remainder sodium bromide, 0.18 mmol active
30 starburst dendrimer), from Example FF, was dissolved in 4.5 ml of deuterium oxide. The resultant pH was 11.5-12. A solution of yttrium acetate was prepared by dissolving yttrium chloride (0.15 g, 0.5 mmol) and sodium acetate (0.41 g, 0.5 mmol) in 1.5 ml of
35 deuterium oxide (2.9 moles of yttrium per mole of dendrimer). Aliquots of 0.5 ml of the yttrium acetate

solution were added to the dendrimer solution and the ^{13}C NMR spectra recorded at 75.5 MHz.

5 The ^{13}C NMR spectrum of yttrium acetate shows
two resonances, 184.7 ppm for the carboxyl carbon and
23.7 ppm for the methyl carbon, compared with 182.1 and
24.1 ppm for sodium acetate, and 177.7 and 20.7 ppm for
acetic acid (Sadtler ^{13}C NMR Standard Spectra).
10 Monitoring the positions of these bands indicates
degree of chelation with the starburst dendrimer. The
most informative signal for the starburst dendrimer
which is indicative of chelation is the $\alpha\text{-CH}_2$ (of the
methylene carboxylate group involved in chelation),
15 which appears at 58.4 ppm in the unchelated dendrimer,
and 63.8 ppm in the chelated dendrimer. Upon chelation
with yttrium, the spin lattice relaxation times of the
time $\alpha\text{-CH}_2$ shortens as expected from $0.24 \pm 0.01\text{s}$ to
0.14 $\pm 0.01\text{s}$, indicative of chelation.
20

Following the addition of 0.5 ml of the yttrium
acetate solution to the starburst dendrimer, all the
yttrium appeared to be chelated by the dendrimer,
25 confirmed by the signals for the acetate being that of
sodium acetate. The same observation was noted for the
addition of a second 0.5 ml aliquot of the yttrium
acetate solution. Upon addition of the third aliquot
of yttrium acetate, not all of the yttrium was observed
30 to be taken up as the starburst chelate, the acetate
carboxyl resonance was observed to shift to 183.8 ppm
indicating that some of the yttrium was associated with
the acetate. The integrated area of the chelated -CH_2
35 groups on the dendrimer increased, indicating that some
of the third mole equivalent of yttrium added was

indeed chelated with the dendrimer. These results indicate that the dendrimer can chelate from 2-3 yttrium ions per dendrimer molecule.

- 5 Example 9: Demonstration of Multiple Chelation of Yttrium by a methylene carboxylate terminated second generation starburst polyamidoamine by trans chelation from yttrium acetate.

10 The same experimental methods were used for this study as were used for Example 8. The starburst polyamidoamine methylene-carboxylate terminated material (0.40g 62.5% active, remainder sodium bromide, 0.12 mmol.) was dissolved in 4-5 ml of deuterium oxide. The resultant pH was 11.5-12, which was lowered to 9.4
15 with 6N HCl prior to the experiment. A solution of yttrium acetate was prepared by dissolving yttrium chloride (0.1125g, .37 mmol.) and sodium acetate (0.0915g, 1.1 mmol.) in 1.5 ml of deuterium oxide, thus
20 every 0.5 ml of solution contains one mole equivalent of metal.

 The first two mole equivalents of yttrium acetate added were fully chelated by the starburst
25 polyamidoamine. On addition of a third mole equivalent of yttrium, precipitation of the product occurred and as such no NMR data could be obtained. The signals which gave the most information about chelation by the starburst dendrimer were those of the two carbons
30 adjacent to the chelating nitrogen. The chemical shifts of these carbons in the unchelated dendrimer occurred at 59.1 ppm for the α -CH₂, and 53.7 ppm for the first methylene carbon of the backbone. Upon
35 chelation these two resonances were observed to shift downfield to 60.8 and 55.1 ppm respectively. The trans chelation shows that two metal ions can be readily

chelated per dendrimer molecule, however upon chelation of some unknown fraction of a third mole equivalent, the product precipitates out of solution.

5 Example 10: Demonstration of Multiple Chelation of ^{90}Y by a methylenecarboxylate terminated second generation starburst polyethyleneimine.

Standard solution of yttrium chloride (3×10^{-2} M, spiked with non-carrier added ^{90}Y) and
10 methylenecarboxylate terminated second generation starburst polyethyleneimine (6×10^{-2} M) were prepared. These were reacted together at various metal:starburst ratios in HEPES buffer. The complex yield was
15 determined by ion exchange chromatography using Sephadex G50 ion exchange beads, eluting with 10% NaCl: NH_4OH , 4:1 at pH 10. Noncomplexed metal is removed on the column, complexed metal elutes. Yields
20 were obtained by comparing the radioactivity eluted with that on the column, using a well counter.

25

30

35

Table V

Chelation of 2.5 Gen. PEI Acetate with ^{90}Y

5

	<u>Vol. Y+3</u>	<u>Vol. PEI</u>	<u>Vol HEPES</u>	<u>M:L Theor.</u>	<u>% Complex</u>	<u>M:L Act.</u>
	5	30	370	0.1	110	0.1
	10	30	360	0.2	101	0.2
10	20	30	350	0.4	95	0.4
	30	35	340	0.5	97	0.5
	30	30	340	0.5	102	0.5
	60	30	310	1.0	99	1.0
15	120	30	250	2.0	100	2.0
	180	30	180	3.0	94	2.8
	250	30	120	4.1	80	3.3
	300	20	80	7.5	44	3.3
20	300	20	70	5.0	40	2.0
	300	20	70	5.0	41	2.0

All volumes in Table V are in microlitres

25 Within the accuracy of the experiments, these results indicate that the 2.5 Gen. starburst PEI acetate can chelate between 2 and 3 metals per polymer giving a soluble complex.

30

35

Example 11: Conjugation of 4-isothiocyanatophenyl methylenecarboxylate terminated third generation starburst polyethyleneimine with IgG monoclonal antibody

5 The isothiocyanate, 10 mg (50 μ moles), from
 Example DD was dissolved in 500 μ l of 3 mM indium
 chloride which had been spiked with radioactive indium-
 111 chloride and the pH was adjusted to 9 with 660 μ l N
 NaOH. Aliquots of whole antibody IgG CC-46 was then
 10 mixed with aliquots of the chelated starburst. The
 mixtures were shaken then left for 18 hours. The
 mixtures were then analyzed by HPLC (column Dupont
 Zorbax Biosphere GF-250; eluent 0.25 M sodium acetate,
 pH 6) and a UV detector at 254 nm and a radioactivity
 15 detector. Results are shown in Table VI.

Table VI
 Starburst-IgG Conjugates

	<u>1</u>	<u>2</u>	<u>3</u>	<u>4</u>
20 IgG solution (μ l)	20	20	20	20
Chelated Starburst solution (μ l)	5	20	50	100
25 % Radioactivity on IgG	6	5	5	3
% IgG conjugated	2	7	17	22

30 Example 12: Conjugation of 4-isothiocyanatophenyl methylenecarboxylate terminated third generation starburst polyethyleneimine with IgG monoclonal antibody

35 The isothiocyanate from Example DD, 4 mg (20 μ moles) was mixed with 200 μ l of 3 mM indium chloride (60 μ moles). A 20 μ l aliquot of the solution was then
 spiked with radioactive indium-111 chloride and the pH
 adjusted to 9 by the addition of 30 μ l 1 N NaOH and 10

5 μ l of 0.1 HCL. The indium chelate was mixed with 150
10 μ l of CC-49 whole antibody IgG, 10 mg/ml in 50 mM HEPES
buffer at pH 9.5. After 18 hours at room temperature
the antibody was recovered by preparative HPLC (column
Dupont Zorbax Biosphere GF 250; eluent 0.25 M sodium
acetate, pH 6); and a UV detector at 254 nm and a
radioactivity detector. The recovered antibody was
concentrated on an Amicon membrane and exchanged into
PBS buffer at pH 7.4. The recovered antibody had
specific activity of approximately 0.5 μ ci/100 μ g.

Example 13: In Vivo localization of ^{111}In labeled
starburst antibody conjugate.

15 The usefulness of the labeled starburst
antibody conjugate prepared in Example 12 was
demonstrated by measuring the uptake of the material by
a human tumor xenograft in an athymic mouse. Female
anthymic mice were inoculation subcutaneously with the
20 human color caicinoma cell line, L-174T (approximately
 4×10^6 cells/animal). Approximately two weeks after
innoculation, each animal was injected via the tail
vein. The mice were sacrificed after 17 and 48 hours
25 (five animals at each time point), the tumor and
selected tissues were excised and weighed, and
radioactivity was measured in a gamma counter. After
17 hours 13.5 percent of the injected dose per gram of
tissue had localized at the tumor. After 48 hours 21.6
30 percent of the injected dose per gram of tissue had
localized at the tumor.

Example 14: Attachment of herbicidal molecules (2,4-D) to the surface of starburst dendrimers.

Third generation PAMAM (initiator core= NH_3) (2.0 g, 0.8 mmole) was dissolved in H_2O (10 ml) and
5 combined with toluene (20 ml). The two-phase system was then stirred and cooled with an ice bath at which time the acid chloride of 2,4-D [2,4-dichlorophenoxy)-acetic acid] (2.4 g, 12 equiv) dissolved in toluene (10 ml) was added dropwise over 30 minutes. When the
10 addition was nearly complete, NaOH (0.5 g, 12.5 mmole, 50% w/w solution) was added and the solution stirred for an additional two hours. The reaction mixture was then evaporated to dryness and the resulting solid
15 residue repeatedly taken up in $\text{CHCl}_3/\text{MeOH}$ (1:1) and filtered. The tan solid was not totally soluble in CHCl_3 and appeared to be insoluble in water; however, the addition of acetone facilitated dissolution. The tan solid was stirred in CHCl_3 for 24 hours and the
20 solution filtered (a sticky tan solid was obtained). After drying over MgSO_4 , the filtrate was concentrated to give a viscous orange oil which solidified on standing. The ^{13}C NMR partial amidation at the surface
25 by 2,4-D is consistent with the association of the 2,4-D to starburst polymer.

Example 15: Inclusion of 2,4-dichlorophenoxyacetic acid (2,4-D) and 2-(acetyloxy)benzoic acid (aspirin) into starburst dendrimers.

30 A widely accepted method for ascertaining whether a "probe molecule" is included in the interior of a micelle is to compare its carbon-13-spin lattice relaxation times (T_1) in a non-micellized versus
35 micellized medium. A substantial decrease in T_1 for the micellized medium is indicative of "probe molecule"

inclusion in the micelle. Since starburst dendrimers are "covalently fixed" analogs of micelles, this T_1 relaxation time technique was used to ascertain the degree/extent to which various herbicide type molecules were associated with starburst polyamidoamines. In the following examples, T_1 values for 2,4-dichlorophenoxyacetic acid (I) and (2,4-D) were determined in solvent ($CDCl_3$) and then compared to T_1 values in $CDCl_3$ at various [I:dendrimer] molar ratios.

Inclusion of 2,4-D into various starburst polyamidoamine dendrimers as a function of generation.

Various half generation (ester terminated, initiated off NH_3) starburst polyamidoamine dendrimers (Generation (Gen) = 0.5, 1.5, 2.5, 3.5, 4.5 and 5.5) were combined with 2,4-dichlorophenoxyacetic acid (I) in $CDCl_3$ to give an acid:tertiary amine ratio of 1:3.5 and molar ratios of acid:dendrimer of 1:86 as shown in Table VII. The relaxation times (T_1) obtained for the various carbon atoms in 2,4-dichlorophenoxyacetic acid and a generation = 3.5 starburst PAMAM dendrimers are shown in Table VIII, both for 1:1 acid/amine ratios and for saturated solutions of 2,4-D.

Table VII

	Gen	(A) Acid/Amine	(B) Acid/Amine	(C) Acid/Total Nitrogen	(D) Molar Ratio (Acid/Star- burst)
5	0.5	1	--	1	1
	1.5	1	1.33	0.57	6
	2.5	1 (3.5)*	1.11 (3.8)*	0.53 (1.8)*	9 (34)*
	3.5	1 (3.0)*	1.05 (3.2)*	0.51 (1.6)*	20 (67)*
10	4.5	1	1.02	0.51	42
	5.5	1	1.01	0.50	86

* represents examples of 2,4-D inclusion into the interior of the dendrimer in amounts greater than stoichiometric.

Table VIII

T_1 's for 2,4-D/G = 3.5 PAMAM Starburst
Inclusion complex: Concentration Effects

		(A)		(B)	
	<u>Carbon</u>	<u>1:1 Acid/Amine</u>		Saturated with	
				<u>2,4-D</u>	
		T_1	$^{13}C^{**}$	T_1	$^{13}C^{**}$
25	1	3.19±.12	(152.73)	3.08±.09	(152.30)
	3	0.34±.01	(128.64)	0.29±.01	(129.62)
	5	0.38±.01	(127.41)	0.32±.01	(127.34)
	2	3.28±.08	(125.79)	2.72±.68	(125.99)
	4	4.58±.16	(123.27)	3.95±.07	(123.16)
	6	0.31±.01	(114.66)	0.28±.01	(114.48)
	CH ₂	0.16±.01	(67.29)	0.146±.003	(66.79)
30	C=O	1.24±.07	(170.12)	-	-

** represents ^{13}C chemical shifts referenced to chloroform at 76.9 ppm..

These data show that larger than stoichiometric amounts of 2,4-dichlorophenoxyacetic acid (i.e., [(I):Gen=3.5 dendrimer]) = 67 can be used without increasing the T_1 in any case in the saturated state (see Columns (A) and (B) in Table VIII). In fact, the relaxation times T_1 (Column (B) are decreased slightly, thus indicating that larger than stoichiometric amounts of 2,4-dichlorophenoxyacetic acid can be included into the interior of the dendrimer. For example, a molar ratio of

[(I):Gen=2.5 dendrimer] = 34 whereas [(I):Gen=3.5 dendrimer] = 67,

(see Column D in Table VII).

Figure 11 is a plot of T_1 values for carbons-3, 5 and 6 in 2,4-dichlorophenoxyacetic acid as a function of dendrimer generation (i.e., 0.5 \rightarrow 5.5). A minimum in T_1 is reached in all cases of generation 2.5 \rightarrow 5.5, thus indicating incorporation in that dendrimer generation range is occurring. Figure 11 also includes T_1 values for 2,4-D in the presence of triethylamine [N(Et)₃] and N(Et)₃ + N-methylacetamide. It can be seen that these values are much larger than for dendrimers G = 1.5 \rightarrow 5.5, thus further supporting molecular incorporation into the dendrimer molecule.

Example 16: Preparation of a product containing fluorescein with a starburst polymer

A sample of 5-carboxyfluorescein (0.996 g) and starburst polyethyleneimine (Gen=2.0; amine terminated, initiated off NH₃) (0.202 g) were mixed in 10 ml of methylene chloride and 5 ml of methanol and allowed to reflux for 10 minutes. Upon filtering, an insoluble

red powder (0.37 g) was obtained (mostly unreacted 5-carboxy fluorescein). From the filtrate was isolated 0.4 g of a brilliant-red solid which exhibited a softening point of 98-103°C and foamed to a brilliant red melt at 175-180°C; NMR spectra (D₂O) of this product were consistent with dendrimer having fluorescein bound to the surface.

Example 17:

10

In a procedure similar to that described in Example 3, starburst polyethyleneimine (Gen=2.0; amine terminated, initiated off NH₃) was reacted with fluorescein isothiocyanate to give a brilliant-red iridescent solid which was suitable for use as a fluorescent labelling reagent.

15

Example 18: Encapsulation of R(+) - Limonene in Polyamidoamine Starburst Dendrimers

20

A 5-50 weight percent solids solution in methanol of starburst - PAMAM dendrimer (M.W. about 175,000; generation = 9.0) is added dropwise to R(+) limonene in methanol until saturated. The solution is stirred at room temperature (about 25°C) for several hours and then devolatilized on a Büchi rotovap at room temperature to give a solid product. Warming at temperatures greater than 80°C gives solvent insoluble products which retain substantial amounts of R(+) limonene in an encapsulated form. These products are excellent prototypes for slow release of (R+)-limonene as a fragrance and deodorizer product.

25

30

35

Example 19: Encapsulation of Heavy Metal Salts in Polyamidoamine starburst Dendrimers

A 5-50 weight percent solids solution in water of starburst PAMAM dendrimer (M.W. about 350,000; generation = 10.0) is stirred as a saturated solution of lead acetate $[\text{Pb}(\text{C}_2\text{H}_3\text{O}_2)_2]$ is added dropwise. The solution is stirred at room temperature (about 25°C) for several hours and then devolatilized on a Büchi rotovap to give solid products. Scanning transmission electromicrograph of these products showed that these heavy metal salts are encapsulated in the interior of the dendrimers. These films containing heavy metal salts are useful as shields for absorbing electromagnetic radiation.

Example 20: Encapsulation of Fluorescein (water soluble) Dye in Polyamidoamine Starburst Dendrimers

A 5-50 weight percent solids solution ($\text{H}_2\text{O}/\text{CH}_3\text{OH}$) of starburst-PAMAM dendrimer (M.W. about 175,000; generation = 9.0) is stirred as fluorescein, disodium salt (Acid Yellow 73, Cl. 45350; Uranine; available from Aldrich (Chemical Co., Milwaukee, WI) is added until saturated. The solution is stirred at room temperature (about 25°C) for several hours and then devolatilized at room temperature to give a colored solid product. These dye encapsulated dendrimers are excellent reference probes for calibrating ultrafiltration membranes.

Example 21: Preparation of dendrimers with terminal fluorescent groups

A. Reaction of Amine Terminated Dendrimer with N-Donxyl Aziridine

A sample (1.5 g, 1.6×10^{-3} mole) of starburst polyethyleneimine (LPEI), $G = 3.0$, terminal groups

(Z) = 12, M.W. = 920) was dissolved in 20 ml of methanol. The solution was stirred and 0.884 g (3.84×10^{-2} mole) of a solution of N-dansyl aziridine (ICN Biomedicals, Costa Mesa, CA) was added dropwise over a period of 20 minutes. The reaction mixture was allowed to stir at room temperature overnight. Removal of solvent under vacuum gave a solid product. NMR and infrared analysis indicated that the product was covalently bonded dansyl groups in the surface of the dendrimer.

B. Reaction of Amine Terminated Dendrimers with Dansyl Chloride

A solution of starburst polyamidoamine (1.0 g, 1.9×10^{-4} mole) (initiated from ammonia, G = 4.0, terminal groups (Z) = 24, M.W. = 5,147) in 30 ml of water was stirred in a 3-neck flask with 80 ml of toluene while a solution of dansyl chloride (1.23 g, 4.5×10^{-3} mole) (5-dimethyl-amino-1-naphthalene-sulfonyl chloride, from Aldrich Chemical Co., Milwaukee WI) in 40 ml of toluene was added dropwise while cooling with ice. Concurrently, a solution of 10% NaOH (13.3 mole, 10% excess) was added to the reaction mixture to give an oily ball. The product was washed with water, dissolved in methanol, and precipitated with diethyl ether to give a solid product. NMR and infrared analysis was consistent with covalently bonded dansyl groups in the dendrimer surface.

WE CLAIM:

1. A starburst conjugate which comprises at least one starburst polymer associated with at least one unit of at least one carried material.

5 2. The conjugate of Claim 1 wherein the starburst polymer is a starburst dendrimer.

 3. The conjugate of Claim 1 wherein the carried material is a pharmaceutical material
10

 4. The conjugate of Claim 1 wherein the carried material is an agricultural material.

 5. The conjugate of Claim 1 wherein at least
15 one of the carried materials is a drug, pesticide, radionuclide, chelant, chelated metal, toxin, antibody, antibody fragment, antigen, signal generator, signal reflector, or signal absorber.

20

25

5 6. The conjugate of Claim 2 wherein there are at least two different carried materials at least one of which is a target director.

10 7. The conjugate of Claim 2 wherein there are at least two different carried materials at least one of which is a target director and at least one of which is a bioactive agent.

15 8. The conjugate of Claim 7 wherein the target director is an entity specific for one or more target receptors and the bioactive agent is a radionuclide, drug, pesticide or toxin.

20 9. The conjugate of Claim 7 or 8 wherein the target director is a polyclonal antibody or fragment thereof.

20 10. The conjugate of Claim 7 or 8 wherein the target director is a monoclonal antibody or fragment thereof.

25 11. The conjugate of Claim 1 wherein the dendrimer contains discontinuities.

30 12. A starburst conjugate of Claim 1 of the formula:



wherein each P represents a dendrimer;

35 x represents an integer of 1 or greater;

each M represents a unit of a carried material, said carried material can be the same carried material or a different carried material;

5 y represents an integer of 1 or greater; and

* indicates that the carried material is associated with the dendrimer.

10 13. The conjugate of Claim 12 wherein M is a drug, pesticide, radionuclide, chelant, toxin, antibody, antibody fragment, antigen, signal generator, signal reflector, or signal absorber.

15 14. The conjugate of Claim 12 wherein x=1 and y=2 or more.

20 15. The starburst conjugate of Claim 12 wherein the molar ratio of any ionic M to P is 0.1-1,000:1.

25 16. The starburst conjugate of Claim 12 wherein the weight ratio of any drug, pesticide or toxin M to P is 0.1-5:1.

30 17. A process for preparing



35 wherein each P represents a dendrimer; x represents an integer of 1 or greater; each M represents a unit of a carried material, said carried material can be the

same carried material or a different carried material; y represents an integer of 1 or greater; and * indicates that the carried material is associated with the dendrimer, which comprises reacting P with M, usually in a suitable solvent, at a temperature which facilitates the association of the carried material (M) with the starburst dendrimer (P).

18. The process of Claim 17 wherein the temperature is from room temperature to reflux.

19. The process of Claim 17 wherein the suitable solvent is water, methanol, ethanol, chloroform, acetonitrile, toluene, dimethylsulfoxide or dimethylformamide.

20. A starburst conjugate of Claim 3 or Claim 4 which also has at least one agricultural or pharmaceutically acceptable diluent or carrier present.

21. The starburst conjugate composition of Claim 20 which also has other active ingredients present.

22. A starburst conjugate of any one of Claim 3 or Claim 4 for use as a diagnostic agent.

23. A starburst conjugate of Claim 3 for use as a therapeutic agent.

24. A method for the delivery of at least one carried agricultural or pharmaceutical material which

comprises administering of at least one starburst conjugate of Claim 3 or Claim 4 containing said material at or near a targeted locus.

5 25. A method for scavenging therapeutic or diagnostic compounds which comprises administering of a bifunctional starburst conjugate containing a target director of Claim 7 which localizes the conjugate to a target locus and a scavenging moiety which can bind a
10 secondarily administered therapeutic or diagnostic compound.

15 26. The method of Claim 25 wherein the scavenging moiety is a chelant, antigen, or antibody.

27. A starburst conjugate of the formula

20
$$[(T)_e - (C')_f]_g * (P)_x * [(C'')_h - (M)_y]_k \quad (III)$$

wherein

25 each C' represents the same or different connecting group;

 each C'' represents the same or different connecting group;

30 g and k each individually represent an integer of 1 or greater;

 f and h each individually represent an integer of 0 or greater;

35

- indicates a covalent bond in instances where a connecting group is present; and

each P represents a dendrimer;

5 x represents an integer of 1 or greater;

T represents a target director;

10 each M represents a unit (for example, a molecule, atom, ion, and/or other basic unit) of a carried pharmaceutical material, said carried pharmaceutical material can be the same carried pharmaceutical material or a different carried material, preferably
15 the carried pharmaceutical material is a bioactive agent;

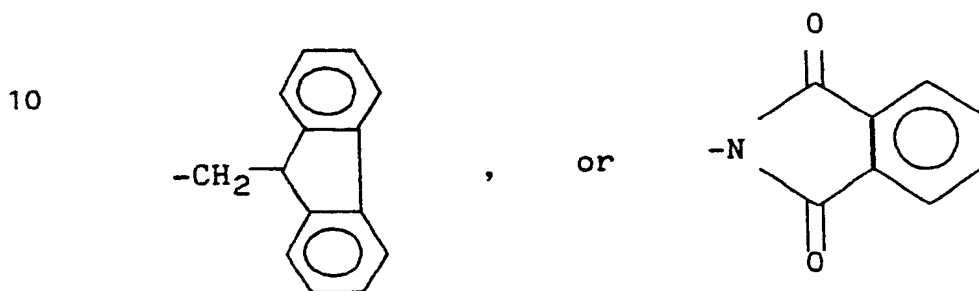
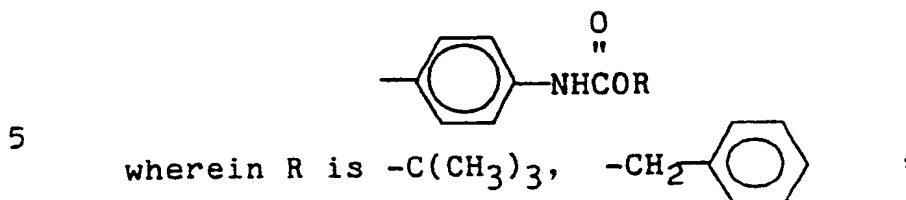
y represents an integer of 1 or greater; and

* indicates that the carried pharmaceutical material is
20 associated with the dendrimer.

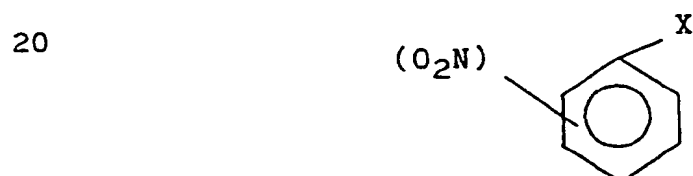
28. A process for preparing a starburst conjugate as defined in Claim 1 which comprises the reaction of P, having reactive moieties, with an
25 aniline moiety, which may have the NH₂ group protected by a carbamate.

29. The process of Claim 28 wherein the carbamate has the formula
30

35



15 30. The process of Claim 28 wherein the P also has a connecting group (handle), C' and/or C'', attached of the formula



25 where n is 1 or 2, and X is F, Cl, Br, I, SO₂Cl, and when n is 1, the NO₂ group is in the para position.

30 31. The process of Claim 30 wherein the connecting group is 4-fluoronitrobenzene.

35

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Neu eingereicht / Newly filed
Nouvellement déposé

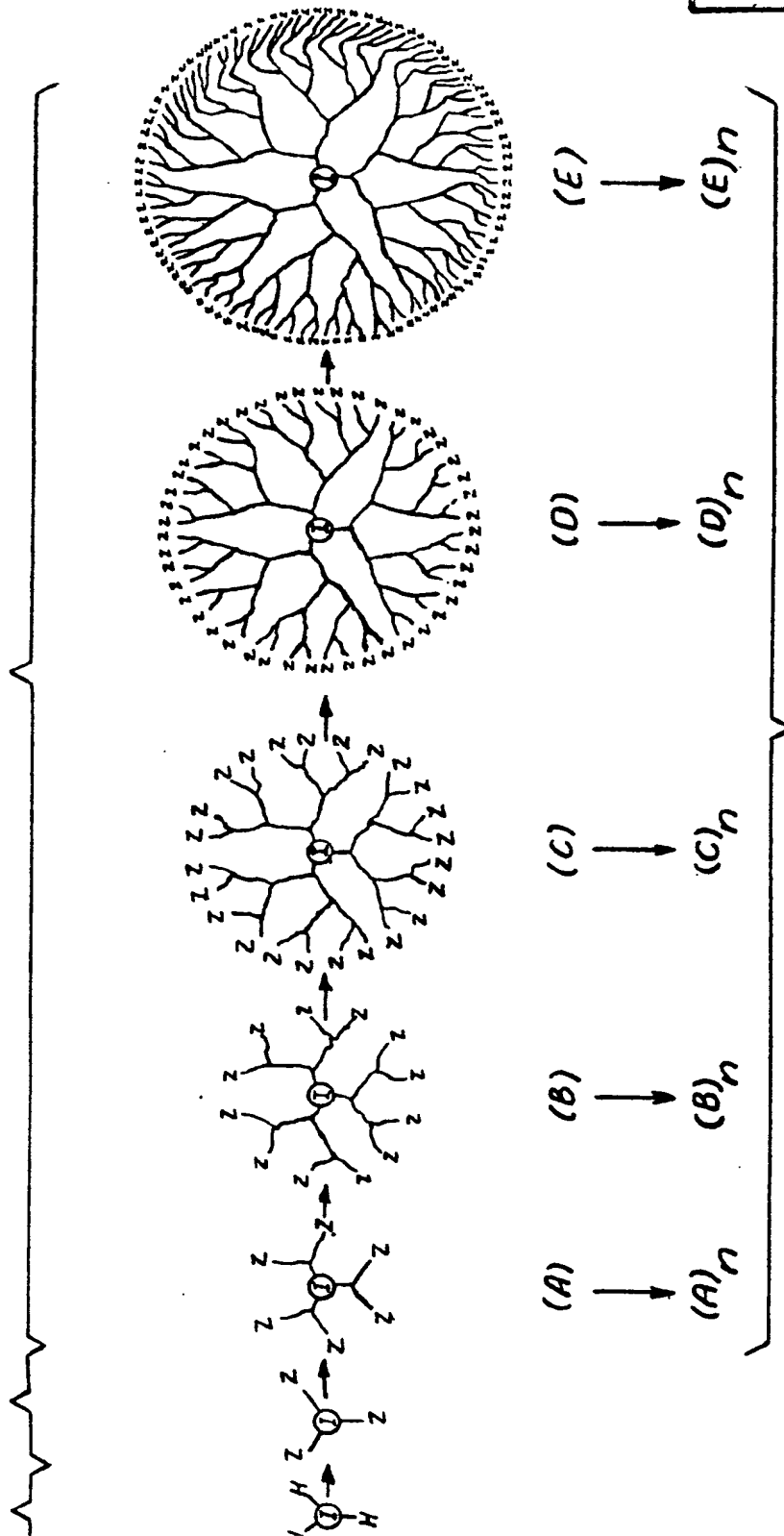
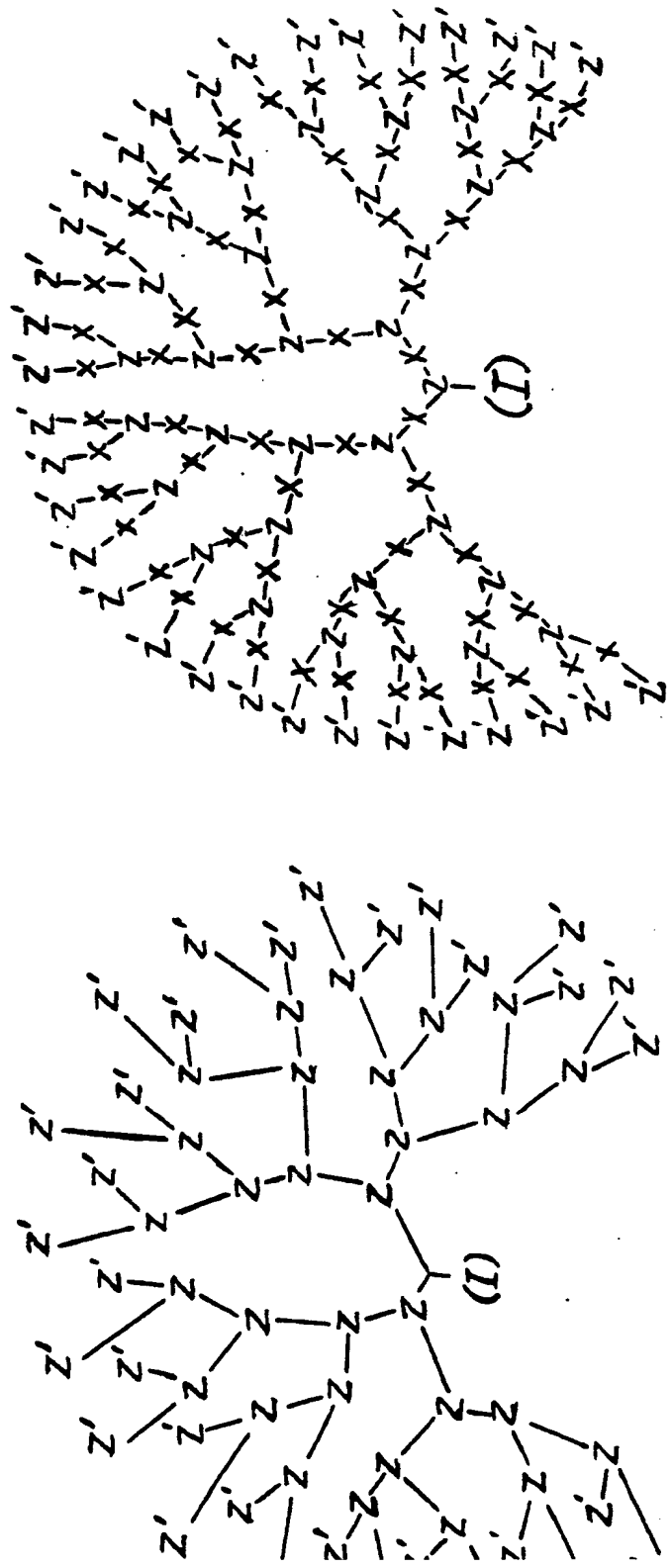


Fig. 1

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0 271 180

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Nouvellement déposé



(B)

(A)

Fig. 2

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Not registered / Breveté
Nouvellement déposé

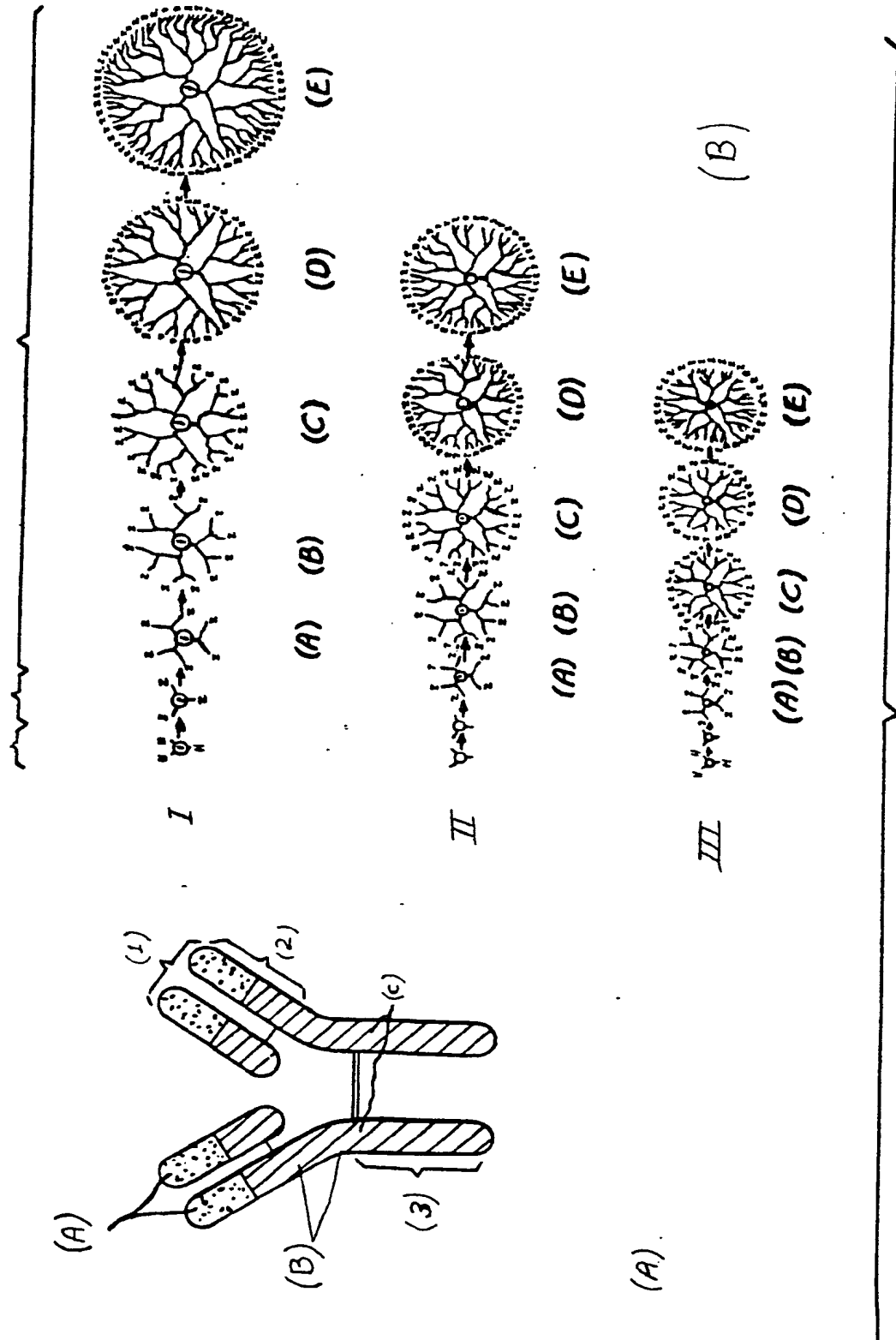
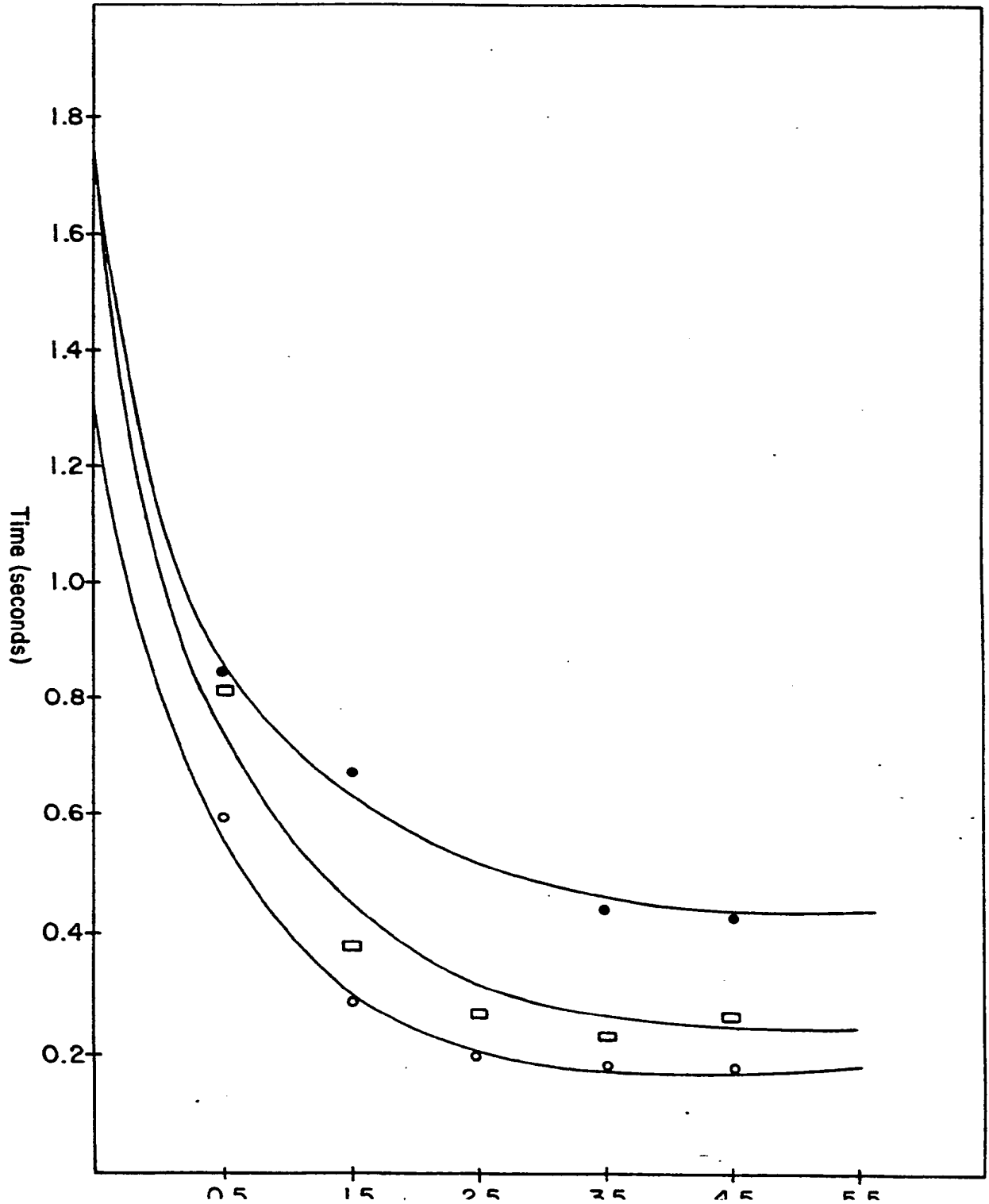


FIG. 3

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Nouvellement déposé

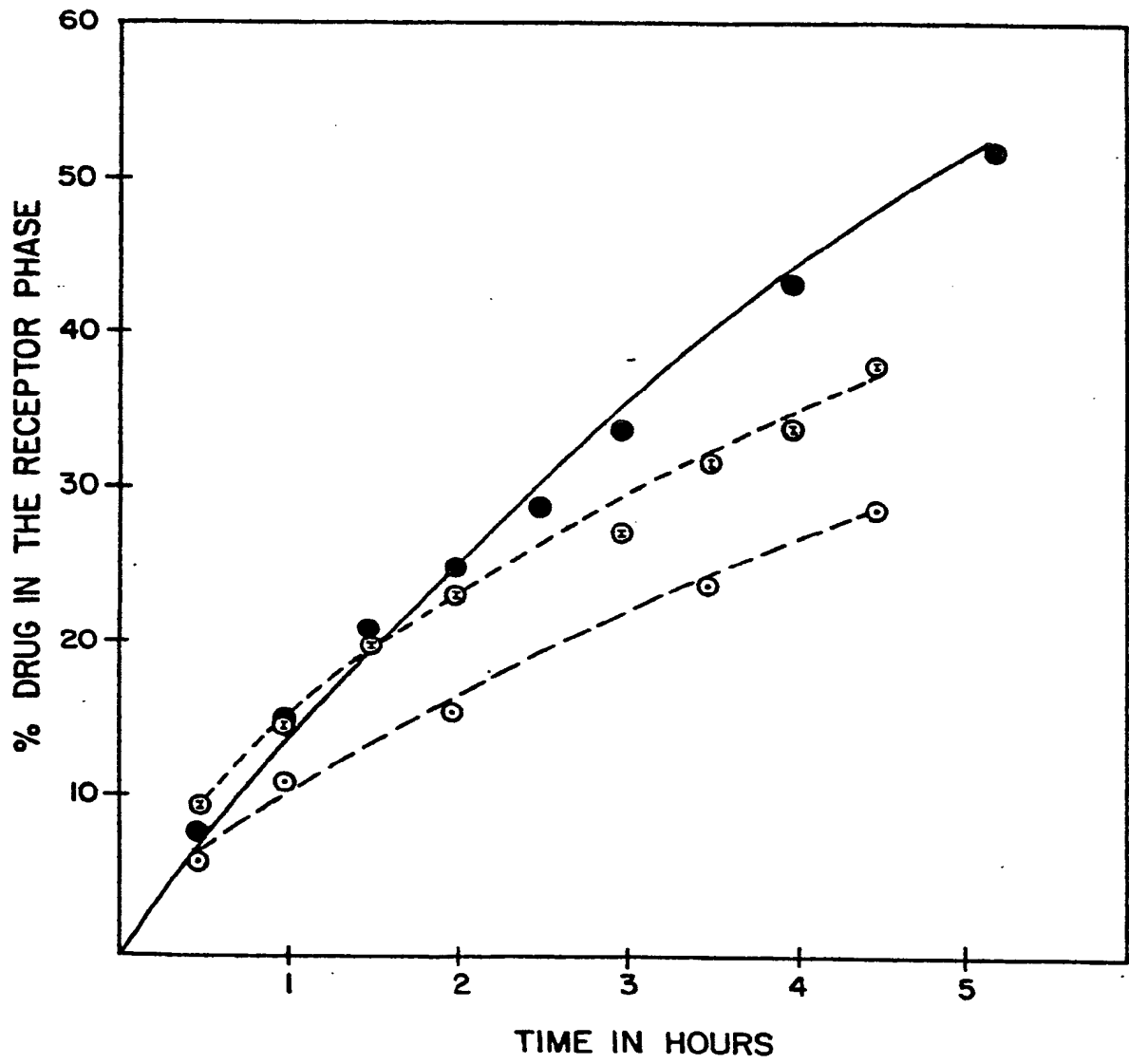
FIG. 4



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Neu eingereicht / Newly filed
Nouvellement déposé

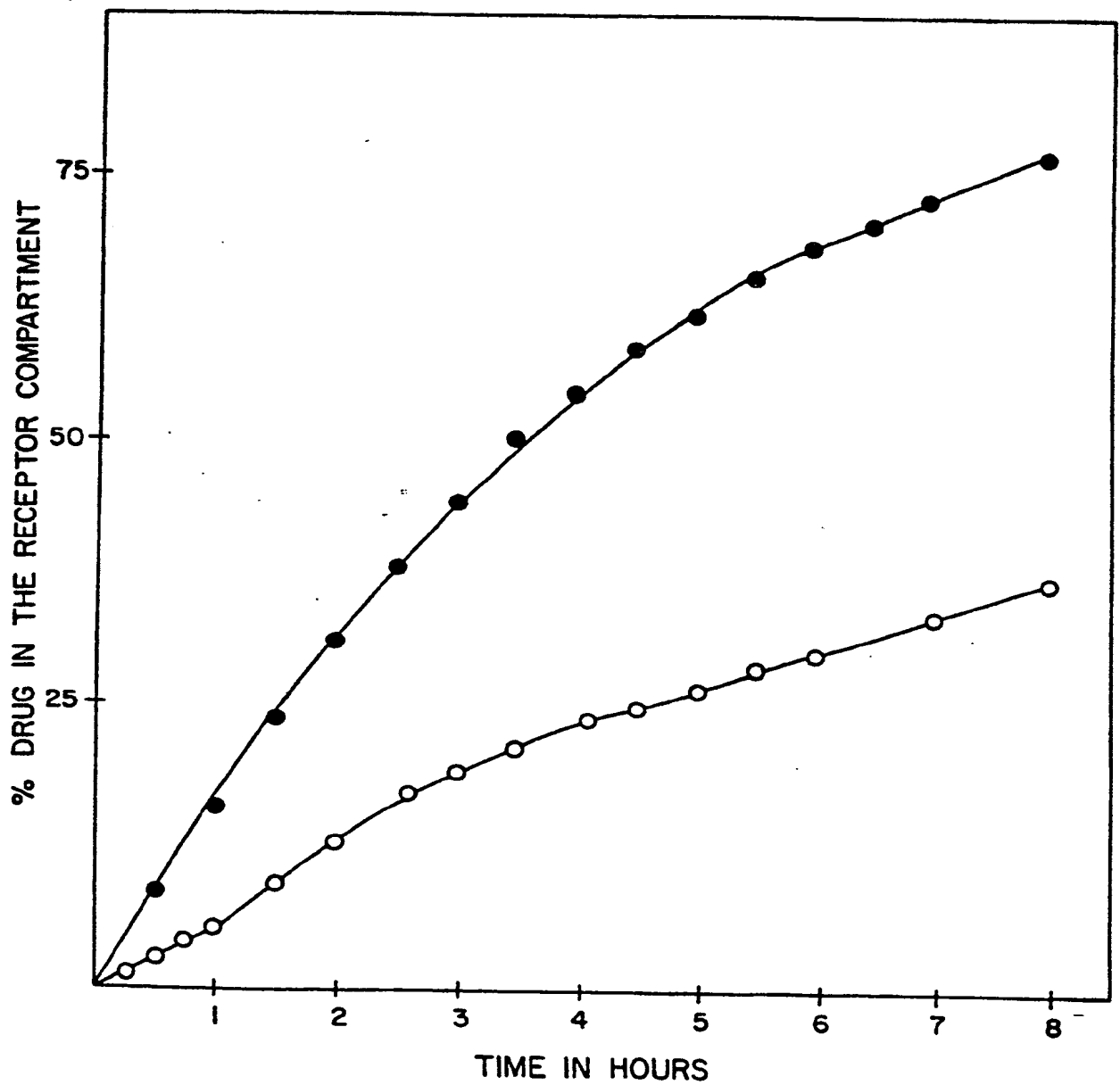
FIG. 5



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Not eingereicht / Newly filed
Nouvellement déposé

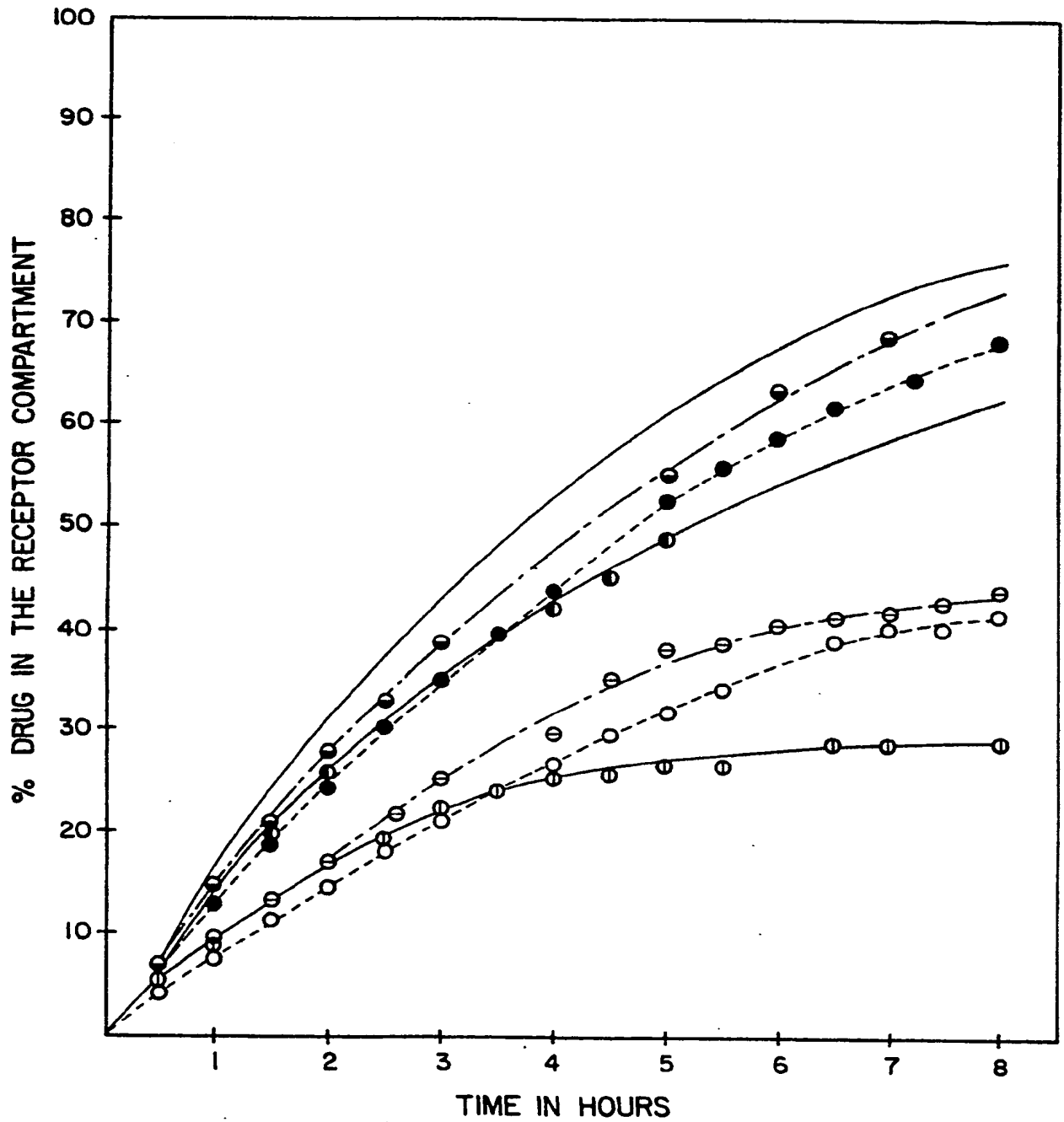
FIG. 6



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FIG. 7



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FIG. 8

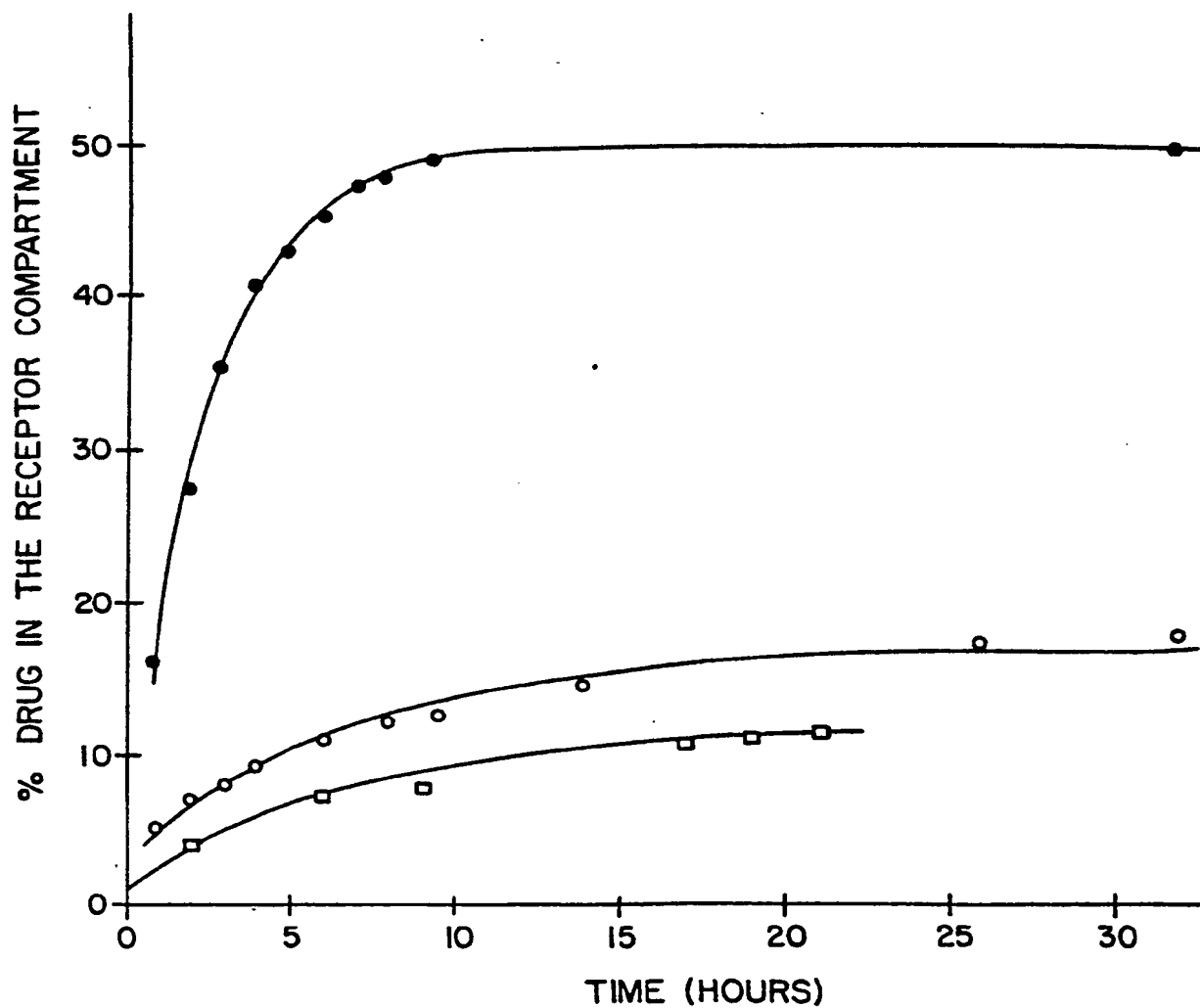
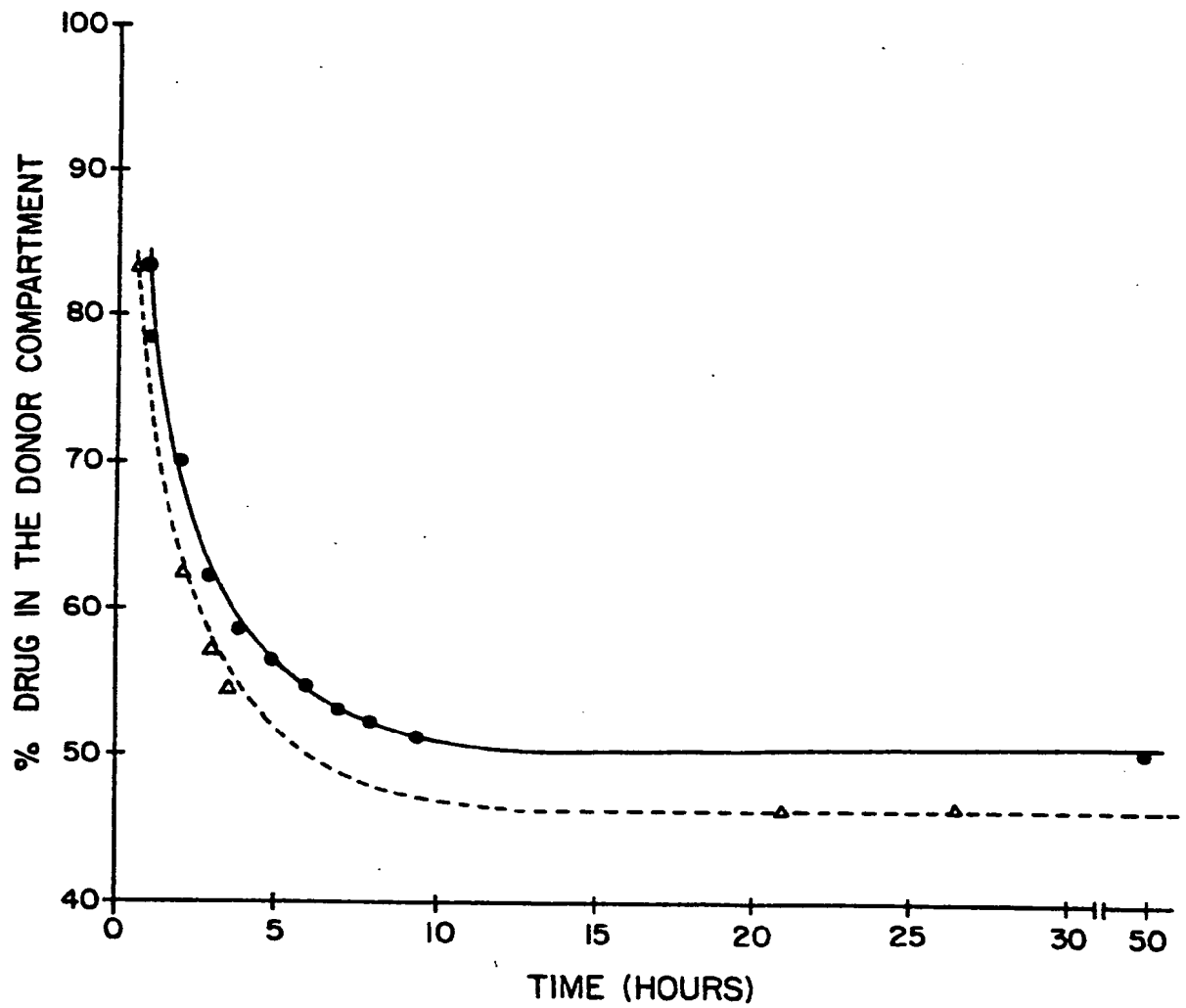


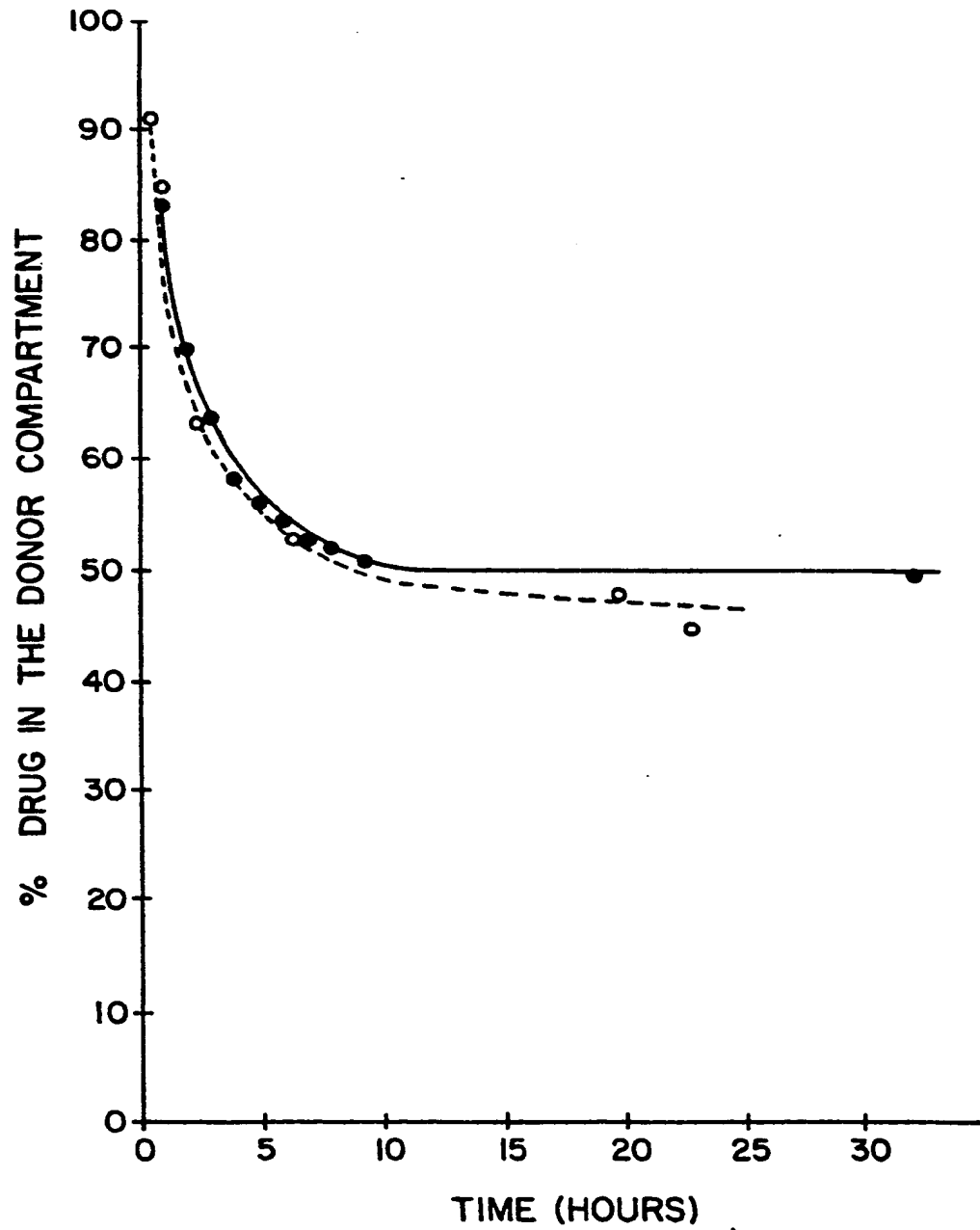
FIG. 9



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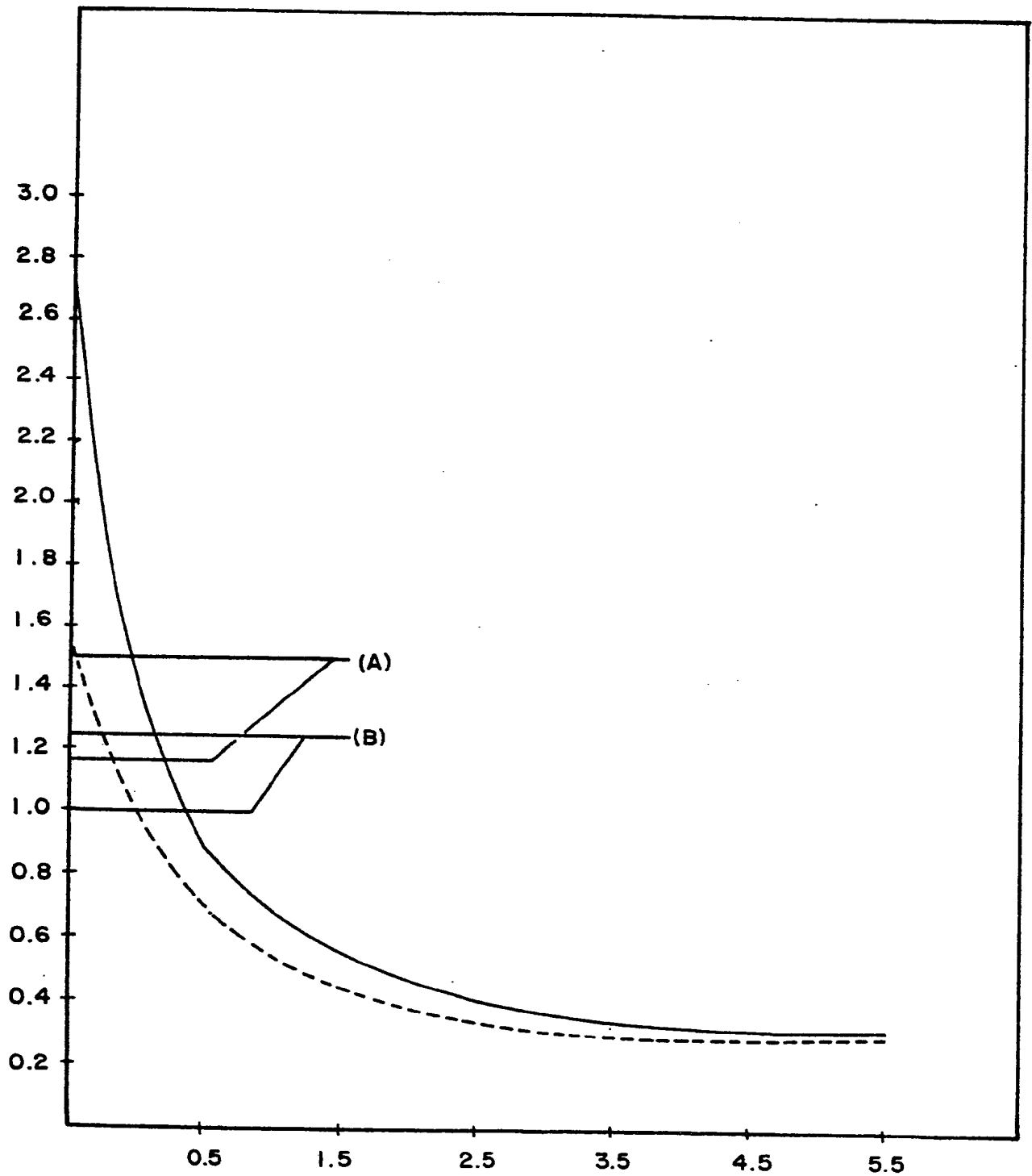
Neu eingereicht / Newly filed
Nouvellement déposé

FIG. 10



Neu eingereicht / Newly filed
Nouvellement déposé

FIG. II





DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.4)
D,X Y	US-A-4 289 872 (R.G. DENKEWALTER) * Abstract, last 3 lines; claim 5; column 4, lines 32-39 * ---	1-26	A 61 K 47/00 A 01 N 25/10
D,Y	US-A-4 472 509 (O.A. GANSOW) * Column 3, lines 11-50; column 6, line 52 - column 7, line 25 * ---	3-26	
D,Y	EP-A-0 115 771 (THE DOW CHEMICAL CO.) * Claims; abstract * ---	1	
P,A	US-A-4 631 337 (D.A. TOMALIA) * Abstract; claims * -----	1	
			TECHNICAL FIELDS SEARCHED (Int. Cl.4)
			A 61 K A 01 N
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 30-11-1987	Examiner BERTE M.J.
CATEGORY OF CITED DOCUMENTS			
X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background		T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons	